Effects of nano, chelated and conventional iron as soil addition on growth, anatomy and early yield of cucumber plants (*cucumus sativus l*).

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ABSTRACT

Two greenhouse experiments in Randomized complete block design with four replications were conducted at Soil, Water and Bioenergy Resources, South Centers, Piketon, OH, Ohio State University, USA, to study the effect of iron forms (conventional Ferric chloride (FeCl₃. 6H₂O),chelated iron(6% Fe) and Nano Iron Oxide (Fe₂O₃, alpha, 99%,)at different doses(0, 50 and 100 mg/kg⁻¹ soil) on growth, anatomy and early yield in cucumber plants (*Cucumus sativusL*.) cv.(Tyria F1) during 2015 and 2016 seasons. Results showed that, the different applied treatments increased plant height, leaf number, leaf area (LA) (cm²), fresh weight and dry weight at60 and 90 days after transplanting during the both seasons of cultivation. Also, anatomical studies were carried out on stems i.e.(stem thickness, xylem and Phloem thickness, and numbers, cambium region thickness, Fiber tissue thickness and pith thickness) and leaves measurements i.e.(plaside and spongy tissue and xylem thickness, number of vessels and phloem at 30 days of plant age in the second season. Data showed positive response to the different applied treatments especially with nano iron at 100 mg/kg⁻¹. In addition, early fruits number/plant, fruit length, fruit diameterand earlyfruit weight per plant increased due to the different applied treatments in the first and second seasons. Results also, showed that soil application of nano iron at 100 mg/kg⁻¹ is the most effective treatment in this respect compared with other tested iron sources and control treatments.

Keywords: nano ferric, chelated, growth, early yield, anatomy.

Introduction

With the development of nanotechnology, nanomaterials are being used in the field of agriculture (Li et al., 2014). Nanomaterials consist of nanometerscale particles with a very small diameter and a large specific surface area. In comparison with traditional materials; nanomaterials have many special functions resulting from the quantum size effect, microscopic quantum tunneling, and dielectric confinement effect (Michal et al., 2005; Hochella et al., 2008), and also have some new functions (Gui et al., 2015). Consequently, such nanomaterials have many potential applications. . In September 2003, the United States Department of Agriculture stated the importance of using nanotechnology in agricultural production; since then, many other countries have increased research efforts in this field (Sanguansri and Augustin, 2006). In addition, many experts have proposed that nanoparticles should be used in the field of soil-plant nutrition to achieve sustainable development of agricultural production with minimal environmental impacts (Sastry et al., 2007). Nowadays, nanotechnology considered a new technical revolution. Therefore, nanomaterials will become the new materials for agricultural development, and represent new ideas and directions for global agricultural production (Chen and Yada, 2011). Iron (Fe) is an essential nutrient for all organisms (Zuo and Zhang, 2011) and its deficiency is widespread among many different crops (Sánchez-Alcalá et al., 2014). Despite of Fe content in soil is usually high, but a large proportion is being fixed to soil particles (Mimmo et al., 2014; Bindraban et al.,2015). Also, Fe is mainly in the form of insoluble Fe3+, especially in high-pH and aerobic soils; therefore, these soils are usually deficient in the available form of Fe2+.Because plants usually absorb Fe2+ from soil, Fe-deficient soils lead to Fe-deficient plants (Kobayashi and Nishizawa, 2012). In plants, Fe participates in many physiological processes including chlorophyll biosynthesis, respiration, and redox reactions. Hence, iron fills many essential roles in plant growth and development, including chlorophyll synthesis, thylakoid synthesis and chloroplast development (Miller et al., 1995).(Mimmo et al., 2014; Ye et al., 2015; Zargar et al., 2015). In this respect, since. Iron (Fe) is a cofactor for approximately 140 enzymes that catalyze unique biochemical reactions (Brittenham, 1994). Iron is required at several steps in the biosynthetic pathways., Fe deficiency not only affects the growth and development of plants, but can also lead to anemia in animals and humans (Li et al., 2014). Therefore, it is important to improve the utilization efficiency of Fe fertilizers. The idea is quite simple: nanodevices can help to deliver the nutrients in the right place at the right time and to reduce the action of external agents leading to losses by degradation, etc. The expected effect should be a reduction in the amount of active chemicals incorporated into the plants and soils leading to a reduced negative impact on the environment. However, many researches and knowledge are needed to accomplish those goals not only for the development and synthesis of nano carriers and nano materials, but also for studying the interactions of such nano devices with the plants and the environment. Indeed, it is necessary to study how plants absorb and uptake nanoparticles, how they move inside the plant, and how they interact with the plant cell. Cucumber is one of the vegetable crops which is very important for the human nutrition and human health due to its high content from fiber, minerals and many other compounds. Foliar application of chelated iron increased vegetative and reproductive growth and fruit quality of cucumber plants(Mohsen 2013). Nano fertilizer has a positive effect on spinach wet weight by increasing growth indexes such as leaf area index, crop growth rate and leaf numbers of treated plants based on the same effect of different nano fertilizer concentrations (Alirezaladanet al., 2012). reported that the epidermis cells of the control were similar in shape and size, while the epidermal cells of the NPtreated leaves became larger in size and reached a maximum size when 3mg/kg⁻¹ Fe₃O₄ nano Particles NPS foliar spray was used. In addition, the thickness of mesophyll tissue, which is specialized photosynthetic tissue that contains chloroplasts in palisade and spongy parenchyma tissue, was greater in Fe₃O₄ treated leaves compared to control leaves. The leaf spray treatment was also more effective than soil treatment, and the air spaces were largest in foliar sprayed leaves. Therefore, foliar spray was the most effective treatment, followed by soil treatment, compared to control leaves. This finding was clear based on the chlorophyll concentration, which was higher in leaves receiving the foliar treatment compared to soil treatment. Therefore, this study aimed to evaluate the effect of different doses of Nanoiron as soil addition on growth, growth

performance, anatomical structure of (stem and leaf)and early fruits yield in cucumber plants in comparison with other iron forms i.e., conventional or chelated ones.

Materials and Methods

Two greenhouse experiments in randomized complete block design (RCBD) with four replications were conducted at Soil, Water and Bioenergy Resources, South Centers, Piketon, OH, Ohio State University, USA to study the effect of iron sources conventional ferric chloride (FeCl₃₋ 6H₂O), chelated iron (6% Fe) and nano Iron oxide (Fe₂O₃, alpha, 99%, 30-100 nm) at doses of 0, 50 and 100 mg/kg⁻¹ soil on growth, anatomical structure and fruit yield of cucumber, (Cucumus sativusL.)(Tyria F1) during 2015-2016 and 2016-2017 seasons. Seeds of cucumber cv.(Tyria F1) (Cucumus sativusL.) imported from LTD. USA company. the seeds were sowing in October 6th cucumber seeds were planted respectively in the growth chamber and were moved from the growth chamber to the green house on November 13thcucumber seedlings were transplanted to the greenhouse in plastic bags (30 cm) the bags size 12 lb (5.44 kg) of soil media promixpx ingredients (Canadian sphagnum peat moss (75-85 %), perlite horticulture grade, vermiculite horticulture grade, dolomitic and cacitic limestone (PH adjuster) wetting agent, starter nutrient. Imported from premier tech horticulture INC.127 South 5th street, suite 300 Quaketown, BA USA. 18951.

All the different designed treatments were added to the soil after 10 days from transplanting in both the first and second seasons. All recommended agricultural managements through plant growth and development were applied in both seasons.

The characteristics of both the irrigation water quality and growing media are presented in Table1.

рН	dsm ⁻¹ Zn	Р	K	Ca	Mg	S	Al	В	Cu	Fe	Mn	Mo	Na	
7.3	0.71	6.5	36	Elem 208	ental co 24	o mposi 20	tion of t 13	0	ation w 1.6	ater (mg 7.7		0.3	12	1.4
4.7	0.8	118	617	Elem 112	ental co 31	o mposi 84		he Grov 3.2		lia chara 2.5	acteristi 3.7	i cs (mg 0.3	g/kg⁻¹) 43	0.7

Table 1. Main characteristics of the Water used for irrigation and plant growth media.

Iron Sources:

Conventional Fe fertilizer (FeCl₃- 6H2O):

Ferric chloride certified A.C.S. (Lump) FeCl₃- 6 H2O F.W. 270.32 limit about 0.002 % P T. imported from Fisher Scientific Company.

Chelated Iron:

EDDHA Chelated Iron 6%: Ethylenediamine-N,N'-bis (2-hydroxyphenylacetic acid) imported from Grow More Company.

Nano iron: Nano iron imported from Nanostructured & Amorphous Materials, Inc. 16840 Clay Road, Suite #113, Houston, TX 77084, USA

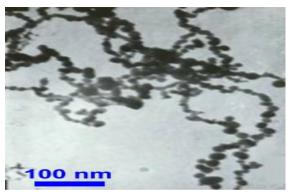


Fig (1) Nano particlesCharacteristics under electronic microscope from Nano structured & Amorphous Materials, Inc. Company.

All iron sources at different doses were added to the soil one time after 10days from transplanting during 2015 and 2016 seasons.

Data Recorded

I- Morphological growth measurements:

Different morphological characteristics of cucumber plants were measured at 60 and 90 days after transplanting. These characteristics involved: Plant height (cm), number of leaves / plant, shoot fresh weight and dry weight (g) / plant and total leaf area /plant (cm²) using Handheld Laser Leaf Area Meter imported from CID Bio-Science,Inc,1554 NE 3rd Avenue Camas, WA USA 98607.The samples of each treatment were dried in oven at 65 °C for 48 hours to take the dry weight .

II- Anatomical studies:

At 30 days after transplanting in the second growing season specimens of stem and leaf were taken from the 2nd apical internode of the stem and from the second apical leaf. Those vegetative specimens were killed and fixed in F.A.A. according to (Sass, 1951). The selected materials were washed in 50 % ethyl alcohol, dehydrated in a normal butyl alchohol series, embedded in paraffin wax of melting point 56°C, section to thickness of 20 micrometer (µm), double stained with saffarine and fast green, cleared in xylene and mounted in Canada balsam (Nassar and El-Sahhar, 1998). Four section treatments were microscopically inspected to detect histological manifestations of noticeable responses resulted from treatments. Count and measure (μ) were taken using a micrometer eye piece.

III-Early Yield and Yield components:

From the harvesting start until reachthree weeks the early yield per plant was calculated (kg/plant),fruit length(cm) , fruit diameter(cm²) and fruits number/plant were measured at the same time. **Statistical analysis:**

Statistical analysis was performed using SAS 9.3 (SAS 2010). Different iron forms were considered as fixed effects. F-protected simple and interactive treatment means were separated by the Tukey's significant difference test, when the ANOVA showed significant effects ($p \le 0.05$) of predictor variables on dependent variables, with a value of p < 0.05 unless otherwise mentioned. Regression and correlation analyses were performed by using SigmaPlot® software.

Results & Discussions

Vegetative Growth parameters:

Data in Table (2) indicates that the different iron forms i.e. the conventional, chelated and nano iron at the different doses 0, 50 and 100 mg/kg⁻¹ significantly increased growth parameters i.e., plant height, leaves numbers, leaf area cm²/plant (LA) at 60 and 90 days after transplanting compared with the control during 2015 and 2016 seasons. The most effective treatment among these iron forms is nano at 100 mg/kg⁻¹ during 2015 and 2016 seasons at the aforementioned different times.

These results are in agreement with those of **Mohamed** *et al.*,(2014) reported that foliar application of wheat plant with nano iron at 0.2.4 and 6 g/l) increased plant height, tillers numbers and number of leaves/plant.

Abo-Sedera *et al.*,(2016) on snap bean and **Abou-Shlell** (2017) on moringa plants reported that foliar application withLithovit (Ca, Mg and Fe nano particles) increased plant height, No. of leaves /plant, No. of branches /plant, fresh and dry weight/ plant and leaf area /plant. **Jayvanth** *et al.*,(2017) found that foliar application of strawberry plants with iron oxide nano particles at 150 mg/l⁻¹ increased plant height, number of leaves and petiole length.,

Data in Table (3) show that nano iron at 100 mg/kg⁻¹ was the most effective treatment on fresh weight and dry weight at 60 and 90 days after transplanting (DAT) during 2015 and 2016 seasons. In context the different applied treatments of iron sources (conventional at 50 and 100, chelated at 50 and 100 and nano at 50 and 100 mg/kg⁻¹) significantly increased the same characteristics at 60 and 90 days after transplanting during 2015 and 2016 seasons. The application of Fe shown significant positive effects, in most cases, on growth measurements and chemical composition of cumin and soybean (**Kobayashi and Nishizawa, 2012**).

Table 2. Effect of Nano, Chelated and Conventional iron on plant height, (cm), leaves number and leaf area (cm²)of cucumber plants at 60 and 90 days after transplanting during 2015 and 2016 seasons.

Charactractics

Treatments				After	60 days			After 90 c	lays				
Iron Source	Dose mg/kg ⁻¹	8		Leaves Number/plant		Leaf Area (cm ²)		Plant Height (cm)		Leaves Number/plant		Leaf Area (cm ²)	
	00	1 st	2 nd	1 st	2^{nd}	1 st	2 nd	1 st	2 nd	1 st	2 nd	1 st	2^{nd}
		Season	Season	Season	Season	Season	Season	Season	Season	Season	Season	Season	Season
CONVFe		206.7b	195.2c	27.4c	30.9c	2461.4c	2393.2c	233.2b	229.1c	35.9a	37.5c	2743.1c	2733.2c
CHELFe		206.8b	197.8b	28.2b	34.1b	2841.7b	2664.9b	233.6b	232.4b	36.0a	39.0b	3037.5b	3103.9b
NANOFe		214.9a	202.0a	28.3a	35.1a	2677.8a	2839.5a	238.5a	237.4a	36.3a	41.9a	3760.3a	3750.2a
	0	205.5y	192.1z	27.7z	29.7z	2326.7z	2258.5z	231.2y	227.8z	32.0z	35.6z	2619.4z	2567.7z
	50	212.8x	200.9y	27.9y	34.8y	2798.8y	2766.9y	234.1x	233.1y	36.7y	41.3y	3423.6y	3403.4y
	100	210.1x	202.0x	28.3x	35.7x	2855.3x	2872.2x	240.1x	237.9x	39.5x	41.5x	3497.9x	3616.1x
							Source x D	ose					
CONVFe	0	204.2	193.4	27.3	29.0	2179.6	2186.3	230.8	227.5	32.5	36.3	2424.8	2412.5
	50	207.1	194.1	27.0	32.0	2513.2	2436.3	235.2	228.0	37.0	38.3	2848.8	2820.5
	100	208.7	198.2	28.0	31.8	2691.3	2557.0	233.7	231.9	38.3	38.0	2955.8	2966.5
CHELFe	0	206.3 ^{ns}	191.1*	28.3 ^{ns}	29.8*	2434.4*	2259.1*	231.1 ^{ns}	228.5*	32.8 ^{ns}	34.5*	2698.5*	2609.7*
	50	208.0	202.5	28.8	35.8	3033.6	2805.9	230.7	232.3	36.5	41.5	3269.9	3309.6
	100	206.2	199.8	27.5	36.8	3057.0	2929.6	239.1	236.3	38.8	41.0	3144.2	3392.4
NANOFe	0	205.9	191.7	27.5	30.3	2366.0	2329.9	231.6	227.6	30.8	36.0	2734.9	2681.0
	50	223.3	206.1	28.0	36.5	2849.6	3058.6	236.3	239.1	36.5	44.3	4152.2	4080.1
	100	215.5	208.1	29.3	38.5	2817.8	3130.1	247.5	245.6	41.5	45.5	4393.8	4489.4

* indicates significant interaction among iron source x dose over time at p<0.05.

¥ Means separated by same lower case letters in each column is not significantly different among iron sources at p<0.05. € Means separated by same lower case letters in each column is not significantly different among iron doses at p<0.05. ns means Non-Significant.

Table 3. Effect of nano, chelated and conventional iron on shoot fresh and dry weight (g) of cucumber plants(*Cucumus satives* L.) at 60 and 90 days after transplanting during 2015 and 2016 seasons.

Characteristics	lu 2010 seasons.		After	60 days			After 90 days					
Treatments Iron Source	Dose mg/kg ⁻¹	Fresh weig	ht (g)/plant	Dry weigl	ht (g)/ plant	Fresh weig	ht (g)/plant	Dry weigh	Dry weight (g)/ plant			
		1 st season	2 nd season									
CONVFe		1190b	1141c	169c	162c	1629b	1607c	248c	249c			
CHELFe		1200b	1198b	198b	201b	1651b	1643b	256b	251b			
NANOFe		1249a	1269a	271a	252a	1779a	1775a	358a	305a			
	0 50 100	1071z 1243y 1325x	1064z 1257y 1287x	168z 228y 243x	148z 224y 243x	1538z 1736y 1784x	1515z 1723y 1787x	217z 277y 367x	224z 288y 293x			
				Source X Do	ose							
CONVFe	0 50 100	1091 1153 1325	1075 1195 1154	157 179 171	143 176 169	1602 1627 1657	1539 1623 1660	232 255 257	234 263 252			
CHELFe	0 50 100	1071* 1262 1267	1066* 1263 1264	180* 199 215	156* 198 250	1508* 1677 1769	1507* 1652 1770	210* 271 286	217* 270 267			
NANOFe	0 50 100	1051 1314 1382	1049 1314 1445	166 305 343	147 299 310	1504 1905 1927	1500 1895 1931	209 306 558	221 332 361			

* indicates significant interaction among iron source x dose over time at p<0.05.

¥ Means separated by same lower case letters in each column is not significantly different among iron sources at p<0.05.

€ Means separated by same lower case letters in each column is not significantly different among iron doses at p<0.05.

II- Anatomical studies:

1-Stem anatomy

Data in Table (4) and Fig (2) indicate that stem diameter, fibers thickness and number, Cambial region thickness, phloem thickness upper and outer, xylem thickness, numberof rows and vessel diameterare increased due to application of iron forms at different doses compared with the control in cucumber plants. The highest valueswere achieved due toapplication of nano iron source at 100 mg/kg⁻¹ dose in different traits. Meanwhile, control treatment gave the lowest value in different stem anatomy parameters. The increase existed in stem diameter with different applied iron forms were preceded with obvious increases in the phloem units that well help the plant to translocate more photosynthates production from sources (leaves and stem) to fruits, thereby, increased in fruits size and number per plant (Table 6).

2-Leaf anatomy

Data in Table (5)and Fig (3) indicate that different iron applied treatments i.e. conventional at 50, 100 mg/kg⁻¹, chelated at 50and 100 mg/kg⁻¹ and nano at 50 and 100 mg/kg⁻¹ increased thickness of leaf midrib, length of large vascular bundle, width of large midrib in the vascular bundle, , phlome thickness, (upper&lower), xylem thickness, and rows number, vessel diameter, plaside and spongy tissue thickness compared with the control treatments in cucumber leaf at 30 days after transplanting in second season.

The increases of the anatomy measurements could be attributed to the beneficial effects of nano iron on increasing the different structures of leaf tissue during plant growth and development, i.e. palisade and spongy tissue thickness, outer and inner phloem, different xylem parts (thickness, numbers and size). The increment of the conductive tissues (xylem &phloem) are also of a great importance because they could be also involved in the interpretation about why vigorous growth and high yielded fruits were existed with different applied treatments specially in case of nano iron at 100 mg/kg⁻¹. The positive alterations in cucumber stem &leaf anatomy treated with different treatments led to vigorous growth and enhancement of flowering and fruit setting of treated plants. Also, it could be noticed that the increases in stem &leaf anatomy measurements were completely reversed upon vegetative and reproductive growth of treated plants. So the, present study revealed increases of xylem tissue, i.e. the route of mineral nutrients and water translocation from roots to leaves and the phloem tissue (outer and inner) i.e., the pathway of different assimilates from leaves to fruits and different sinks in cucumber plants., thereby, improvements of translocation events directly could be considered as a direct reason for increment the final fruit yield.

In general, the stimulatory effects of applied iron forms on the anatomy features of treated plants could be attributed to the effect cambium activity. The increment of cambium activity could mainly attributed to the increase of endogenous hormones level especially cytokinins and auxins, (**Sotiropoulos** *et al.*,2002).,thereby the plant yield and yield component will increase (Tabe 6).

These results are in agreement with Agamy (2004)onfennel plants and Abbas (2013) on dill plants, showed that all rates of foliar application of Fe at 0, 50, 100 and 150 mg/l significantly increased cortex, thickness, number and thickness of vascular bundles and vascular units (diameter of stem) while pith thickness decreased with the same foliar application. Abou-Shlell (2017) reported that foliar application of moringa plants with Lithovit(nano particles) at 500mg/l increased anatomical measurements in stem and leaf i.e. stem diameter, phloem and xylem thickness and vessel diameter.

III- Early yield and its components (quality)

The results in Table (6) and Fig (2) indicate that fruit number/plant, fruit length(cm) and fruit early yield kg/plant significantly increased with application of nano iron at 100 mg/kg⁻¹ by 1.5 to 3 times over the control during 2015 and 2016 seasons. While conventional and chelated forms of iron at 50 and 100 mg/kg⁻¹ increased fruit number /plant, fruit length (cm) and fruit yield kg/plant during first and second seasons.

Early fruit number and yield /plant were in parallel to the applied dose of each iron source. Also, the highest early fruit yield /plant was existed with nano iron at 100 mg/kg⁻¹ (8.51 kg) followed by chelated at 100 mg/kg⁻¹ (5.45 kg)and conventional at 100 mg/kg⁻¹ (4.11 kg), while the control value was (3.29 kg), during first season. **Shankramma** *et al.*, (2016) reported that soil addition of nano iron improve tomato yield and quality.

In this respect the obtained increases in cucumber fruit number, length and cucumber yield it augmented plants ability to harvest more light energy by delivering carbon nanotubes into chloroplast, and also carbon nanotubes could serve as artificial antennae that allow chloroplast to capture wavelengths of light which is not in their normal range, such as ultraviolet, green, and nearinfrared(**Cossins 2014**). Also, increased in plant growth measurements (Table 2) and biomass production (Table 3), thereby increased cucumber fruits number and size and cucumber yield production and quality.

Table 4. Effect of nano, chelated and conventional iron on stem anatomy measurements microns (µ)of cucumber plants at 30 days after transplanting in the second cultivation	m
season.	

Measurements micron (µ) Treatments			stem anatomy at 30 days after transplanting									
Treatments			Fibers			Phloem Tl	nickness µ					
Iron source	Dose mg/kg ⁻¹	stem diameter µ	Thickness µ	Num.	 Cambial region thickness μ 	upper	lower	Thicknes µ	No. of rows	Vessel diameter µ		
Cor	ntrol	7283	80	3	33	301	291	370	3	201		
CON	50	8673	95	5	43	310	302	570	3	231		
CON.	100	8538	99	7	51	352	312	550	5	212		
CUEL	50	8852	101	9	95	320	501	591	4	311		
CHEL.	100	9233	105	10	110	315	521	579	5	331		
NANO	50	9434	109	12	130	456	478	632	6	318		
NANO	100	9972	110	18	150	569	577	615	7	415		

N Treatmen	Measurements micron (μ) ts	Thickness of leaf	Length of large midrib vascular	Width of large midrib	Phloem th	ickness. (µ)		Xylem tissue		Plaside tissue	Spongy Tissue
Iron Source	Dose mg/kg ⁻¹	midrib (µ)	¹⁾ bundle (μ)	vascular – bundle(µ)	upper	lower	Xylem tissue thick(µ)	No. of xylem rows	Vessel diameter (µ)	– thick.(μ)	thick.(µ)
C	ontrol	2780	752	315	310	131	310	4	74	112	236
CONV.	50	2876	835	366	280	205	350	5	91	115	315
CONV.	100	3054	982	387	333	215	434	5	112	136	320
CUEI	50	3125	1140	395	363	324	453	6	123	158	341
CHEL.	100	3168	1194	412	351	330	513	6	162	178	381
Nano	50	3289	1287	455	372	352	563	6	168	189	410
	100	3470	1388	562	415	391	582	7	192	226	439

Table 5. Effect of nano, chelated and conventional iron on leaf anatomy measurements micron (µ) of cucumber plantsat 30 days after transplanting during the second cultivation season.

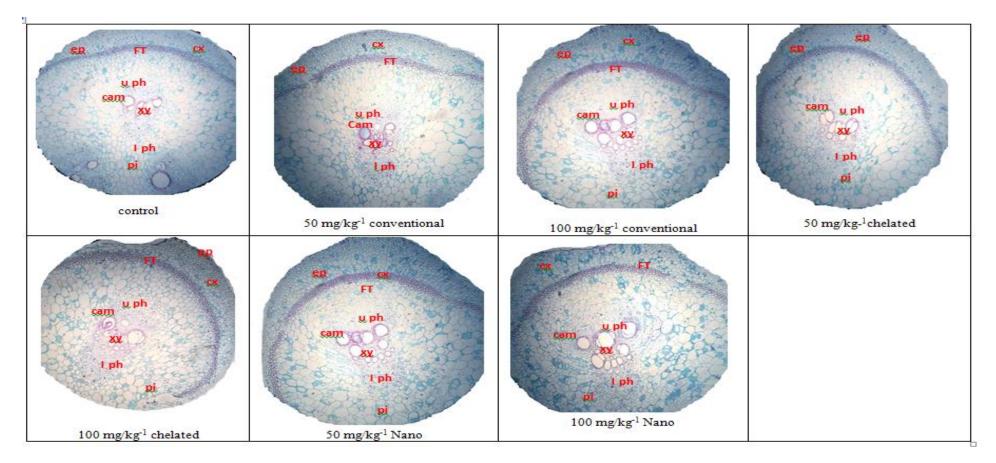


Fig. (2): Transverse sections (X = 24) through 2^{nd} internode of the main stem of cucumber at 30 days after transplanting as affected by the different applied treatments. ep= Epidermis cx= Cortex u ph= upperPhloem tissue I ph= lowerphloem cam=Cambium xy= Xylem tissuepi= Pith

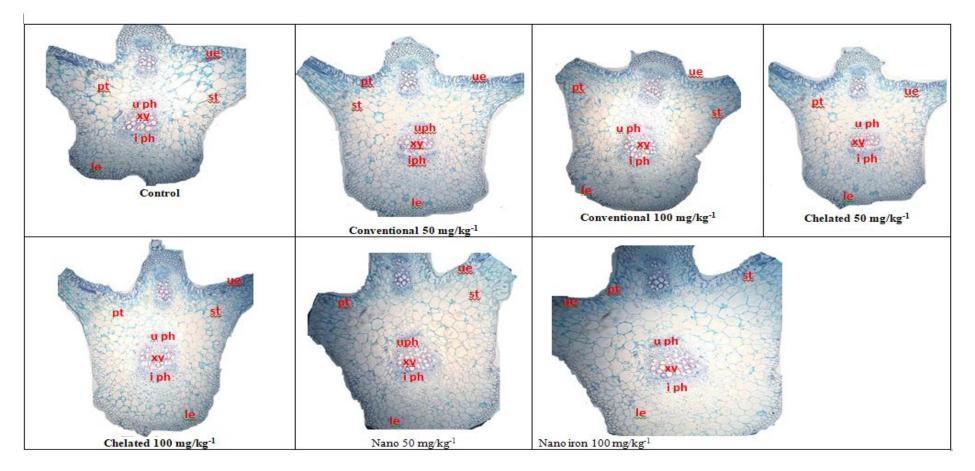


Fig. (3): Transverse sections (X = 24) through 2^{nd} apical leaf of cucumber plants at 30 days after transplanting as affected by the different applied treatments. ue= Upper epidermis pt= Palisade tissue st= Spongy tissue le= Lower epidermis u ph= upper Phloem tissue I ph = lower phloem tissue xy= Xylem tissue

Chara	ctractics									
		Early Fru	iit Number./plant	Frui	t length(cm)	Fruit I	Diameter (cm)	Early Fruit Yield (kg/plant)		
Treatments										
Iron Source	Dose mg/kg ⁻¹	1^{St}	2^{nd}	1^{St}	2 nd	1^{St}	2 nd	1^{St}	2 nd	
		season	season	season	season	season	Season	season	season	
CONVFe		5.00c [¥]	3.75c	27.74b	25.68c	5.58b	5.28c	2.66c	2.27b	
CHELFe		8.50 b	5.00b	32.74b	31.88b	6.23b	6.15b	3.66b	3.27b	
NANOFe		9.50a	6.25a	36.40a	38.94a	7.15a	8.39a	6.16a	5.82a	
0		8.33z€	7.50y	28.78y	28.68y	5.00z	5.72z	3.29z	3.15y	
50		9.66y	8.75y	30.86y	31.67y	7.17y	7.27y	4.99y	4.66y	
100		10.66x	11.00x	34.24x	39.85x	9.15x	9.12x	6.11x	6.55x	
		Source X D	ose							
CONV. 0		3.15*	4.00*	25.68*	28.24*	5.11*	5.24*	2.55*	2.66*	
50		3.52	3.98	28.58	29.34	5.36	5.74	3.15	3.85	
100		4.21	4.80	28.42	31.45	5.98	6.11	4.11	4.25	
CHEL. 0		4.25	4.50	30.35	30.35	5.29	5.47	3.58	3.94	
50		9.50	7.50	27.89	32.97	5.88	6.97	4.36	4.66	
100		9.00	8.25	33.58	33.58	6.87	7.65	5.45	5.08	
NANO 0		6.25	3.50	23.44	28.27	5.36	5.85	4.94	5.88	
50		10.00	11.75	38.94	41.61	8.12	8.46	7.37	6.27	
100		11.50	13.75	48.11	50.57	9.19	10.25	8.51	8.34	

Table 6. Effect of nano, chelated and conventional iron on early yield and yield characteristics of cucumber plants (*Cucumas satives L.*) during 2015 and 2016 seasons.

¥ Means separated by same lower case letters in each column is not significantly different among iron sources at p<0.05.

€ Means separated by same lower case letters in each column is not significantly different among iron doses at p<0.05.

* indicates significant interaction among iron source x dose over time at p<0.05.

Conclusion

Soil application of nano iron at 50 and 100 mg/kg⁻¹ improved plant growth characteristics, shoot fresh and dry weight, stem and leaves anatomical features, early fruit numbers, fruit length and early fruits yield in cucumber plants (*Cucumus sativus*)under greenhouse conditions.

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