



Evaluation of Bioactive Components and Antioxidant Properties of Two Chia Seeds (*Salvia Hispanica L.*)

Kamel, E. B. **; Frahat F.A.Foda*, Abd El-Aleem, I.M.* and Ali, M.A. M. **

* Agric. Biochem. Dept., Fac. Agric., Benha Univ., Egypt.

** Food Technol. Res. Inst., Agric. Res. Center, Giza, Egypt.

Corresponding author: farhat.fouda@fagr.bu.edu.eg

Abstract

The aim of this investigation was to evaluate black chia seed (*Salvia hispanica L.*) as a source of bioactive components comparing with white chia seeds. Also, antioxidant activities, identification of phenolic and flavonoids compounds, fatty acids composition and amino acids of chia seeds content were evaluated. The results showed slight differences in proximate composition, raw black and white chia seeds to have high nutritional content in the form of protein (25.29 – 24.30%), fibre (32.94-31.75%), fat (32.35-30.54%) and ash content (6.05-6.93%), respectively. Concerning to germinated chia seeds, data showed that protein content of white chia seed increased significantly 26.94% compared with raw white chia seeds 24.30%. While, fat and fiber contents were found to be decreased significantly as a result of germination process. Fermented chia seeds showed higher protein content 28.10 and 27.55% for black and white chia seeds, relative to raw chia seeds. Total phenol content was increased a significantly in germinated black seeds (263.51±28.44 mg GAE/100g) relative to raw seeds (166.76±4.13mg GAE/100g). While, fermented seeds showed significant increment in total phenols content for both chia seeds the values were 216.25±4.75 and 200.74±3.64 mg GAE/100g when compared with raw sample. The antioxidant activity as determined by DPPH and ABTS methods was relatively higher in raw black chia seeds than raw white chia seeds. Besides, chia seeds oil was found a high content of linolenic acid (ω 3) and (ω 6) content with a value of 59.25- 58.54% and 20.50-20.17% in black and white chia seeds oil, respectively. Both chia seeds had nearly similar essential amino acids.

Keyword: Chia seeds, germination, fermentation, antioxidant activity, Fatty acids.

Introduction

In recent years, with increased health awareness globally as a result of an increase in the number of non-communicable diseases, the demand for a healthier lifestyle through the consumption of foods that prevent and control these non-communicable diseases has increased (Berner *et al.*, 2014; Chadare *et al.*, 2019). While, all foods are functional with some properties, functional foods have become one of the fastest growing sectors of the food industry with the growing awareness of people about deadly diseases such as cancer, diabetes and heart disease (Hasler and Brown, 2009).

Chia (*Salvia hispanica L.*, family: *Lamiaceae*) was an herbaceous plant used for commercial and medical purposes. Originally, it was grown in tropical and subtropical climates but at present, it is grown all over the world as important food

supplements. Nowadays, dry chia seeds, as important edible seeds, are added to many food-stuffs like fruit juices, smoothies, milk, yogurt, salads, soups, and baked products due to their nutritional properties (Dick *et al.* 2015; Kulczynski *et al.*, 2019). The dry chia seeds contain high contents of essential fatty acids, carbohydrates, minerals, proteins, dietary fibers, and bioactive phenolic compounds (Oliveira-Alves *et al.*, 2017). Chia seeds are generally very small, oval-shaped, 2 mm long, 1 to 1.5 mm wide, and less than 1 mm thick (Mohd Ali *et al.*, 2012; Das 2018; Grancieri *et al.*, 2019). The color of the seed varies from black, grey, or black spotted to white. Moreover, the plant itself can produce 500 to 600 kg seed/acre under appropriate agronomic conditions (Ullah *et al.*, 2016).

Some recent studies reported that the dietary intake of dry chia seeds could protect against obesity-related diseases, oxidative stress, and cardiovascular diseases (Poudyal *et al.*, 2012; Marineli *et al.*, 2015).

Through a simple and inexpensive germination process several edible seeds could be grown in a short time with improved nutritional values and functional properties. During germination, the main macromolecules such as carbohydrates, proteins and fatty acids are degraded into simple/free forms and reduced the anti-nutritional and indigestible factors (Shi *et al.*, 2010; Aguilera *et al.*, 2013). Furthermore, the germination process increased the bioactive-phenolic compounds and many bioactivities in some edible seeds (Gan *et al.*, 2016; Abdel-Aty *et al.*, 2019).

Knez-Hrnčić *et al.* (2018) reported, that a marginal difference between black and white chia seeds that most consider them equal. Nutritional values are similar protein content in black chia seeds was 26.9% and fiber content was 32.6%. In white chia seeds, the protein content was found to be 26.5% and the fiber content was 32.4%. A slight difference was found only in morphology white seeds were larger, thicker, and broader compared to black seeds.

Several studies have been performed on the usage of chia seeds in the food industry can be used in different shapes: whole, ground, in the form of flour, oil, and gel. Also, can be added or mixed into biscuits, pasta, cereals, snacks, and cakes as supplements (Das 2018). Chia gel may be used as substitutes for oil or eggs in baked products. It was shown that chia oil can replace 25% of the egg in cakes (Kulczyński *et al.*, 2019). The nutritional value of butter can be increased by mixing it with chia oil in a proportion from 6.5% to 25%, when the concentration of ω -3 fatty acid in chia fortified butter increases from 4.17% to 16.74% (Ullah *et al.*, 2016). The aim of this work is to evaluate the chemical composition, phenolic and flavonoids components, fatty acids and amino acids contents of two chia seeds genotypes (black and white).

Material and Methods

Chia seeds (*Salvia hispanica*) used in this study were purchased from the local market. All the utilized chemical materials (solvents, mineral salts, *etc.*...) were purchased from El Gomhoryia, El Allamyia and El Nasr Chemical companies, Egypt and the solvents were purified before using. Chemicals, solvents and all standard materials which were used for fractionation and identification by HPLC, purchased from Sigma/Aldrich Chemical Company, USA. 2, 2-diphenyl-1-picrylhydrazyl (DPPH), 2,2-azinobis (3-ethylbenzo-thiazoline-6-sulfonic acid) (ABTS) and Folin-Ciocalteu reagent were obtained from LOBA chemie, from Win. Lab. Laboratory chemicals reagents. All other chemicals used were of analytical reagent grade.

Preparation of raw chia seeds materials

Raw chia seeds were milled in a Laboratory mill (IKA-Laboratechnik, Janke and Kunkel Type: MFC, Germany) to obtain a whole meal flour and kept at -20°C until analysis.

Germination of chia seeds

Germination conditions were established on the basis of preliminary assays according to the method described by (Beltrán-Orozco *et al.*, 2020).

Fermentation of chia seeds

Fermentation condition were established on the basis of preliminary assays according to the method described by (Tamime *et al.* 1997).

Proximal composition

Using de-fatted chia flour, crude protein was calculated by the micro/Kjeldahl method, and moisture, ash and fiber contents were determined by using standard methods (AOAC, 2000). The Soxhlet extraction method was used to measure total fat content. Carbohydrate content was calculated by difference as follows equation: -
Total carbohydrate = 100 - (Moisture + Ash + Crude protein + Crude fat) according to A.O.A.C (2016).

Determination of minerals

Perkin Elmer (Model 3300, USA) Atomic Absorption Spectrophotometer was used to determine these minerals. Such as zinc, iron, calcium, manganese, copper, potassium, phosphorus, sodium and magnesium contents by using methods according to A.O.A.C (2016).

Determination of total phenolic and total flavonoids contents

Total phenolics were determined as described by Singleton and Rossi (1965). Gallic acid was chosen as a standard to prepare the standard curve. Total phenolics was expressed as mg gallic acid equivalent/100g sample dry weight basis. While, total flavonoids were determined according to the method of Zhishen *et al.* (1999). Catechin served as standard compound was used for preparing the calibration curve. Total flavonoids were calculated as mg catechin equivalent (CAE)/100g on dry weight basis.

Identification of phenolic acids and flavonoids compounds by HPLC

The phenolic acids and flavonoids compounds of the samples under investigation were determined according to the method described by Hakkinen *et al.* (1998) and Mattila *et al.* (2000).

Determination of antioxidant activity

The antioxidant activity of chia seeds methanol extracts was determined based on the radical scavenging ability in reacting with a stable DPPH free radical according to Brand-Williams *et al.* (1995). While, the ABTS assay was carried out according to Re *et al.* (1999).

Fatty acid compositions and Identification of the amino acid content

The fatty acid compositions of chia seeds oil were analyzed by Agilent HP 6890 capillary gas chromatography according to the method described by Glass (1971). While, amino acids content of the sample were determined using amino acid analyzer (Biochrom 30) according to Duranti and Cerletti, (1979). Amino acids were expressed as g /100g protein on dry weight basis.

Results and Discussion

The chemical compositions of raw, germinated and fermented chia seeds genotypes (black and white) are measured the obtained results are tabulated in Table (1). From these data showed that raw black seeds had a higher level of protein 25.29% and fat 32.35 compared with raw white chia seeds 24.30 and 30.54%, respectively. In addition, crude fiber and ash contents were also high, they showed nearly the same levels for both chia seeds genotypes (32.94 and 6.05%) for black chia seeds and (31.75 and 6.93%) for raw white chia seeds. The predominant carbohydrate portion (30-34%) of chia seeds is dietary fibre, which cover 100% of the adult population's daily requirements (Motyka *et al.*, 2021)

Concerning to germinated chia seeds, data showed that protein content of white chia seed increased significantly 26.94% compared with raw white chia seeds 24.30%. While, fat and fiber contents were found

to be decreased significantly as a result of germination process. The reason for the decreased fat content is due to its consumption in the germination process to obtain energy. Slight changes occurred in ash content but fiber content decreased in both chia seed genotypes. At the same time total carbohydrates increased significantly by in germinated seeds to 12.70 and 17.04% for germinated black and white chia seeds, respectively.

On the other hand, fermented chia seeds showed higher protein content 28.10 and 27.55% for black and white chia seeds, relative to raw chia seeds. Same trend was observed in fiber content. It recorded 32.67 and 33.55% for fermented black and white chia seeds. From the above-mentioned, the data showed that total carbohydrate slightly decreased compared with germinated seeds. In general germination and fermentation treatments improved the nutritional value of the chia seeds.

These results are in agreement with Suri *et al.* (2016); Ullah *et al.*, (2016); Knez-Hrnčič *et al.*, (2018) and Otondi *et al.*, (2020) they found that Chia seeds had a moisture 6.35%, protein 19.96%, carbohydrate 32.53%, fat 36.61% and ash 4.98%, reported that protein content ranged 15–25% from chia seeds, our value is between this limit. Content of fat of chia seed was of 32.35 and 30.54 for black and white chia seeds. The difference of protein content and oil content could be connected to the difference in climatic conditions, agronomic practices, fertilization regimes, irrigation practices, etc. (Ayerza, 2011).

Table 1. Chemical composition of chia seeds components (%) g/100g dry weight

Samples	%Moisture	%Protein	%Fat	%Fiber	%Ash	%T.C
BR	7.64±0.21 ^a	25.29±0.42 ^c	32.35±0.66 ^a	32.94±0.22 ^{ab}	6.05±0.11 ^b	3.12±1.09 ^c
WR	6.95±0.30 ^a	24.30±0.23 ^d	30.54±0.69 ^a	31.75±1.01 ^b	6.93±0.34 ^a	6.03±1.12 ^c
BG	6.32±0.37 ^b	27.38±0.23 ^{ab}	23.04±1.28 ^c	29.61±0.32 ^c	6.42±0.24 ^{ab}	12.70±1.98 ^b
WG	4.18±0.05 ^c	26.94±0.23 ^b	19.44±0.45 ^d	28.56±0.27 ^c	7.28±0.42 ^a	17.04±0.05 ^a
BF	3.88±0.61 ^c	28.10±0.31 ^a	25.88±0.47 ^b	32.67±0.11 ^{ab}	6.00±0.02 ^b	10.98±0.03 ^b
WF	3.54±0.02 ^c	27.55±0.20 ^{ab}	24.05±0.86 ^{bc}	33.55±0.36 ^a	6.74±0.57 ^{ab}	11.32±1.49 ^b
L.S.D at 5%	0.80	0.83	1.92	1.17	0.83	2.92

TC= total carbohydrates, BR= raw black seeds, WR= raw white seeds, BG= germinated black seeds, WG= germinated white seeds, BF= fermented black seeds and WF= fermented white seeds.

The minerals content of chia seeds (raw, germinated and fermented) are measured and the results are presented in Table (4). The obtained results showed that chia seeds contained appreciable quantities of minerals. They considered a good source of minerals containing calcium, potassium, phosphorus, magnesium, zinc, iron and selenium in relatively higher amounts while, manganese, sodium and copper in smaller quantities. Potassium was found to be abundant mineral in the both raw chia seeds genotypes (black and white) 1667.5 and 1537.5mg/100g, respectively.

While, calcium was found to be the second element in chia seeds, white chia seeds had the highest amount was 879.0mg/100g compared with 609.7 mg/100g for raw black chia seeds. Magnesium element content was 350.85 and 353.1mg/100g in raw black and white chia seeds respectively. Data revealed that both black and white chia seeds had a higher iron level 11.2 and 10.55mg/100g for raw black and white seeds. Also, chia seeds had high zinc level 7.9 and 6.45 mg/100g in black and white seeds. Data revealed that there were

increment in calcium, zinc, iron and copper for germinated and fermented samples.

Ding *et al.*, (2018) reported that the main minerals in the chia seed are magnesium, calcium and potassium as well as iron, zinc, manganese and selenium. da Silva *et al.*, (2017) reported the chia seeds was a highlight due to its concentration of iron, zinc, calcium, manganese, potassium and phosphorus are noted wherein the minerals, calcium (631 mg.100 g⁻¹),

potassium (407 mg.100 g⁻¹), magnesium (335 mg.100 g⁻¹), iron (7.72 mg.100 g⁻¹) and zinc (4.58 mg.100 g⁻¹). These results are in agreement with those reported by Ullah *et al.*, (2016). Kulczyński *et al.*, (2019) they mentioned that chia seeds supply many minerals, with phosphorus of (860–919 mg/100 g), calcium (456–631 mg/100 g), potassium (407–726 mg/100g) and magnesium (335–449 mg/100 g) found in greatest amounts.

Table 2. Content of individual macroelements and microelements (mg/100 g dry seeds) in the chia seeds (*S. hispanica*)

Samples	Minerals content mg/100g dry seeds									
	P	Ca	K	Mg	Na	Fe	Mn	Zn	Cu	Se
BR	160.25	609.7	1667.5	350.85	26.15	11.2	5.05	7.9	0.7	21.65
WR	192.2	879.8	1537.5	353.1	36.7	10.55	7.15	6.45	1.3	20.6
BG	168.65	722.4	2402.5	353.2	91.85	11.1	5.4	9	1.25	20.55
WG	190.2	842.85	1040	356.6	101.4	10.5	6.65	6.75	1.5	18.6
BF	167.25	744.2	817.5	348	520	12.8	4.8	8.55	2.3	15.75
WF	186.35	888.9	1215	355.65	543.55	12.25	6.45	7.05	2.35	14.05

BR= raw black seeds, WR= raw white seeds, BG= germinated black seeds, WG= germinated white seeds, BF= fermented black seeds and WF= fermented white seeds.

Total phenols, total flavonoids and their antioxidant activities: -

Total phenols, flavonoids content and antioxidant activity were measured in raw, germinated and fermented chia seeds and the obtained data are presented in Table (3). Results cleared that total phenols of raw black and white chia seeds were found to be 166.76 ± 4.13 and 182.31 ± 13.35 mg GAE/100g, respectively. Total phenol content was increased significantly in germinated black seeds (263.51 ± 28.44 mg GAE/100g) relative to raw seeds. While, fermented seeds showed significant increment in total phenols content for both chia seeds the values were 216.25 ± 4.75 and 200.74 ± 3.64 mg GAE/100g when compared with raw sample.

In the same Table (3) data showed that total flavonoids in raw black and white chia seeds was 66.85 ± 5.35 and 56.96 ± 5.93 mg CE/100g, respectively. On the other hand, germinated chia seeds, total flavonoids were increased in both genotypes of chia seeds and the values were 70.29 ± 5 and 62.67 ± 6.78 mg CE/100g when compared with raw seeds. Germination probably induced the synthesis of phenolic compounds and also caused the release of these compounds from the food matrix of chia seeds. While, fermented seeds showed decreased significantly in total flavonoids content for black and white chia seeds the amount were 56.56 ± 5.11 and 54.44 ± 2.06 mg CE/100g were compared with raw sample.

The values obtained in this study are similar to those found by other researchers. Reyes-Caudillo *et al.* (2008) reported an average amount of phenolic compounds of 75.7 and 88.1 mg GAE/100 g in chia

seeds from Jalisco and Sinaloa, respectively, whereas Martínez-Cruz and Paredes-López (2014) found an average of 164 mg GAE/100 g in seeds from Colima. On the other hand, Gómez-Favela *et al.*, (2017) reported an amount of 190.8 mg GAE/100 g in chia seeds and observed an increase of 47.4% in total phenolics after 156 h of germination at 21°C. Also, Beltrán-Orozco *et al.* (2020) reported the amount of phenolic compounds was 97.7 mg GAE/100 g in ungerminated seeds, but this value increased 3-fold after 4 days of germination.

The antioxidant activities of chia seeds were investigated using the DPPH and ABTS assays and the obtained values were found to be 86.23 and 77.58 % for raw black chia seeds, respectively. While, raw white chia seeds recorded 85.69 and 76.94 %, respectively. There were non-significant changes between the two chia seeds genotypes. The DPPH values of black germinated and fermented seeds showed non-significant changes relative to raw samples. Same trend was observed in ABTS values for black seeds. While, germinated and fermented white seeds showed a significant decrement in DPPH and ABTS, the values were 59.84 and 52.13%, respectively.

The same results were obtained by other authors such as Alacantara *et al.* (2019) investigated antioxidant activity by the DPPH method it was 10.10 to 380.53 μ mol TE/g extract. Grancieri *et al.* (2019) stated in their research that to investigate the specific antioxidant activity and Beltrán-Orozco *et al.* (2020) they found that the initial antioxidant activities of chia seeds were 77.7 and 41.1 mmol ET/g (dw)

determined by ABTS and DPPH methods, respectively. These values increased by 105.1%, 101.6%, and 87.7%, respectively, after 4 days of germination. Also, **Gómez- Favela *et al.* (2017)** observed an increase of 77.2% and 96.7% in phenolic compounds and antioxidant activity (ABTS method), respectively, after 157 h of germination at 21 °C. The high content of phenolic compounds of chia seeds was strongly related to their ABTS and DPPH radical scavenging activity as

well as to their reducing power. The antioxidant activity was also positively correlated with the content of total flavonoids and ascorbic acid which indicates that all these compounds contributed to the antioxidant capacity of chia seeds. They also found germination has also proved to increase the polyphenol content and the antioxidant capacity of different grains (**Álvarez-Jubete *et al.*, 2010**) and (**Kim *et al.*, 2012**).

Table 3. Total phenols, total flavonoids, DPPH and ABTS in raw, germinated and fermented chia seeds.

Samples	Total Phenols mg GAE/100g	Total Flavonoids mg CE/100g	DPPH %	ABTS %
BR	166.76±4.13 ^d	66.85±5.35 ^a	87.17±1.24 ^a	77.58± 0 ^a
WR	182.31±13.35 ^{cd}	56.96±5.93 ^b	81.48±3.04 ^a	76.94±0.40 ^a
BG	263.53±28.44 ^a	70.29±5.00 ^a	88.81±0.24 ^a	77.76±0.98 ^a
WG	191.03±10.86 ^{bcd}	62.67±6.78 ^{ab}	66.82±9.00 ^b	59.80±5.06 ^b
BF	216.25±4.75 ^b	56.56±5.11 ^b	60.67±1.53 ^b	74.14±4.20 ^a
WF	200.74±3.64 ^{bc}	54.44±2.06 ^b	64.07±0.78 ^b	52.13±1.67 ^c
L.S.D at 5%	24.71	9.33	7.06	4.99

BR= raw black seeds, WR= raw white seeds, BG= germinated black seeds, WG= germinated white seeds, BF= fermented black seeds and WF= fermented white seeds.

Identification of phenolic acids and flavonoids compounds in chia seeds by HPLC: -

Phenolic compounds in chia seeds extracted was identified by HPLC and the obtained results are presented in Table (4). From these data the raw black chia seeds phenolic extract was contained eleven phenolic compounds with concentrations ranging from 0.07 to 6.32mg/100g. Ellagic acid concentration was 6.32mg/100g followed by catechin (5.36 mg/100g), chlorogenic (2.75mg/100g), caffeine (2.13mg/100g) and caffeic acid (1.52mg/100g), respectively. In the raw white phenolic extract, eight phenolic compounds were identified with concentrations ranging from 0.08 to 9.17mg/100g, the main phenolic compounds in both raw chia seeds were catechin and ellagic. New phenolic compound (ferulic) was recognized in black

germinated seeds extract, while there new phenolic (chlorogenic, caffeine and ferulic) were recognized in germinated white seeds extract. Ellagic acid concentrations increased in germinated black and white seeds (20.57 and 12.46mg/100g) compared with raw seeds (6.32 and 5.33mg/100g), respectively. Ellagic acid concentrations increased in both fermented chia genotypes extracts. Black chia extract had higher concentration (32.4mg/100g) than that in fermented white chia seeds extract. In fermented black chia seeds, most of phenolic compounds increased pyrogallol, catechin, 4-amino benzoic, caffeic, vanillic, coumaric acid. Same trend was observed in fermented white chia seeds extract. In addition, coumarin considered new phenolic compound recognized in fermented white chia extract.

Table 4. HPLC of phenolic compounds in chia seeds (mg/100 g dry seeds)

Phenolic acids(mg/100g)	BR	WR	BG	WG	BF	WF
Gallic	0.15	0.23	0.32	0.21	2.75	0.22
pyrogallol	2.45	2.58	1.27	1.33	6.94	2.87
4-Amino-benzoic	0.07	0.08	0.29	0.17	0.6	0.24
Catechin	5.36	9.17	nd	1.88	11.43	2.82
Chlorogenic	2.75	nd	1.58	1.53	4.62	2.93
P-OH- benzoic	1.01	1.28	0.82	0.86	0.77	1.15
Caffeic	1.52	0.89	0.42	0.51	5.47	0.8
Vanillic	0.23	1.33	0.21	0.89	2.29	2.53
Caffeine	2.13	nd	1.46	1.03	2.62	1.17
Ferulic	nd	nd	21.44	10.93	nd	9.02
Ellagic	6.32	5.33	20.57	12.46	32.4	9.56
Coumarin	0.16	nd	1.07	0.6	0.85	0.46

BR= raw black seeds, WR= raw white seeds, BG= germinated black seeds, WG= germinated white seeds, BF= fermented black seeds and WF= fermented white seeds.

These results are slightly difference with those obtained by **Coelho and Salas-Mellado (2014)** mentioned, Chlorogenic acid (4.68 µg/g), Protocatechuic acid ethyl ester (0.74 µg/g), Ferulic acid trace and Quercetin (0.17 µg/g). While, **Abdel-Aty *et al.*, (2019)** mentioned that Pyrogallol, gallic acid, and coumaric acid were also raised 65, 62, and 36 times, respectively, while apigenin was only found in fermented garden cress seeds extract over unfermented extract. But, **Ritthibut *et al.*, (2021)** reported that ferulic and coumaric acids increased 63 and 20 times in fermented rice bran, respectively, compared to unfermented rice bran. Also, caffeic acid was found only in fermented rice bran and not in unfermented rice bran. Also, **Motyka *et al.*, (2022)** reported rosmarinic acid is the dominant phenolic acid with the highest concentration (0.927 mg/g), followed by protocatechuic acid (0.747 mg/g), caffeic acid (0.027 mg/g), and gallic acid (0.012 mg/g).

On the other hand, the flavonoids content (mg/100g) in raw, germinated and fermented black and white chia seeds are estimated and presented in Table (5). From these results flavonoids showed differences between chia seeds genotypes with different

compounds (i.e., Rutin, Naringin, Quercitrin, Apigenin-7-glucose, Quercetin, Naringenin, Kaemp-3-(2-p-comaroyl) glucose and Kaempferol). Naringin was the major compound detected, with values of 10.76 and 17.95mg/100g in raw black and white chia seeds, respectively, followed by Rosmarinic acid (1.98 and 4.13mg/100g) and Rutin 1.28 and 2.1mg/100g, respectively. Apigenin was found in raw black chia seeds while in raw white seeds was not recognized. Flavonoids content increased in germinated chia seeds genotypes extract Rutin compound which disappeared in white chia seeds. Highly increment was observed in Naringin content (12.41mg/100g) for fermented black chia seeds extract compared with raw seeds (10.76mg/100g) while that compound disappeared in fermented white chia seeds extract. In addition, quercitrin increased in both fermented chia seeds genotypes (black and white). In white fermented chia seeds, Quercetin increased (1.96mg/100g) compared with raw seeds (0.8mg/100g). Also, Kaempferol increased in fermented black and white chia (1.87 and 1.57mg/100g), respectively. New flavonoids compound Apigenin was restarted in germinated and fermented white chia 0.45 and 0.44mg/100g, respectively.

Table 5. HPLC of total flavonoids compounds in chia seeds (mg/100 g dry seeds)

Flavonoids components	BR	WR	BG	WG	BF	WF
Rutin	1.28	2.10	19.53	nd	nd	6.49
Naringin	10.76	17.95	nd	6.35	12.41	nd
Rosmarinic	1.98	4.13	22.44	12.91	31.48	9.51
Quercetrin	0.23	0.20	0.2	1.67	4.92	0.91
Apigenin-7-glucose	0.09	0.15	0.15	1	nd	nd
Quercetin	0.80	0.73	0.73	0.85	nd	1.96
Naringenin	0.09	0.07	0.07	0.24	0.41	0.85
Kaemp.3-(2-p-comaroyl) glucose	0.80	0.86	0.86	2.75	12.23	9.97
Kampferol	0.20	0.14	0.14	0.48	1.87	1.57
Apigenin	0.13	nd	nd	0.45	0.33	0.44

BR= raw black seeds, WR= raw white seeds, BG= germinated black seeds, WG= germinated white seeds, BF= fermented black seeds and WF= fermented white seeds.

These values were lower than those reported by **Martínez-Cruz and Paredes-López (2014)** and **Rahman *et al.* (2017)** for non-defatted and defatted chia seeds. **Jin *et al.* (2012)** found Kaempferol 0.013 µg/g), Kaempferol 3-O-glucoside 0.029 µg/g), Epicatechin 0.029 µg/g), Rutin 0.22 µg/g), *p*-Coumaric acid 0.24 µg/g) and Apigenin 0.005 µg/g). **Pellegrini *et al.*, (2018)** mentioned seven polyphenolic compounds were detected in both chia and defatted chia seed samples. Rosmarinic acid was the major compound detected and quantified, with values of 653.98 and 669.88 µg/g in non-defatted and defatted chia seeds, respectively, followed by quercetin.

Fatty acids content of chia seeds oil: -

Fatty acid content for black and white chia seeds are determined and the results are shown in Table (6). Data cleared that chia seeds have been considered a good source of healthy poly unsaturated fatty acids (PUFA). Linoleic acid C18:2 (ω6) content in black chia seeds was similar to white chia seeds (20.50 and 20.17%), respectively. Also, linolenic acid C18:3 (ω3) content in black and white seeds was 59.25 and 58.54% of total fatty acids, respectively. Palmitic acid for black and white chia seeds was 7.5 and 7.77%. Stearic acid C18:0 in white chia seeds was higher than that in black chia seeds (4.05 and 2.83%), respectively. While, oleic acid content in white seed was higher than black seeds (7.98 and 6.71%), respectively. In addition, the n6: n3 ratio was 0.35. **Ixtaina *et al.*, (2010)** reported that n-3: n-6 ratio was higher than 3.5 might reduce cholesterol

levels and improve the plasma lipid. These results are in agreement with their obtained (Di Marco *et al.*, 2020; Enes *et al.*, 2020 and Ghafoor *et al.*, 2020) they reported that chia seeds oil has been contain an excessive number of fatty acids, especially polyunsaturated fatty acids (PUFA), which consist of more than 60% α -linoleic acid and more than 20% linoleic acid.

While, Carrillo *et al.* (2018) reported the composition of omega 3 as 54.08%, the content of omega 6 as 18.69%, and omega 9 as 10.24%. (Das, 2018; Knez-Hrnčič and Ivanovski, 2019 and Kuznetcova *et al.*, 2020) they found that the oil

obtained from the seeds of *S. hispanica* accounts for 30–33% of fatty acids. Also, the ratio of omega-6 to omega-3 fatty acids in chia seeds is very favorable (around 0.30–0.35). Monounsaturated fatty acids, which belong to the group of omega-9 fatty acids, constitute about 10% of the fatty acid pool in chia seeds, with oleic acid being dominant. Saturated fatty acids constitute the remaining 10% of the fatty acid pool, and the dominant ones are palmitic and stearic acid (Coelho and Salas-Mellado, 2014). Rajaram (2014) reported that α -linolenic acid, which is derived from plant origin, has the potential to reduce cardiovascular disease, fracture, and diabetes Type 2.

Table 6. Comparison of fatty acid contents profile of two chia seeds genotypes

Fatty acids (% of total fat content)	Black chia seeds	White chia seeds
palmitic acid C16:0	7.5	7.77
stearic acid C18:0	2.83	4.05
oleic acid C18:1	6.71	7.98
linoleic acid C18:2 n-6	20.5	20.17
linolenic acid C18:3 n-3	59.25	58.54
Saturated fatty acids (SFA)	10.33	11.82
polyunsaturated fatty acids (PUFA)	86.46	86.69
Fatty acids ratios		
n-6: n-3	0.35	0.35

Amino acids content of chia seeds: -

The amino acids content of black and white chia seeds were determined by using amino acid analyzer and the obtained results are presented in Table (7). From these data, the chia seeds contain 18 out of 22 amino acids, of which 10 are essential which contribute significantly to human health. These amino acids can be arranged according to their concentration in the following decreasing order, arginine, leucine, phenylalanine, valine, lysine, methionine, threonine, isoleucine, histidine and tryptophan, respectively. Among non-essential amino acids, glutamic acid found at high concentration followed by aspartic acid, alanine, glycine, serine, proline, tyrosine and cystine. Generally, glutamic acid considered an important amino acid in the diet. It is able to modulate immunoregulatory response and enhances athletic performance.

Also, arginine plays role in preventing heart diseases (Timilsena *et al.*, 2017).

These results are increment with these reported by (Olivos-Lugo *et al.*, 2010) found to originate from essential amino acids such as leucine, isoleucine and valine, and high amounts of glutamic acid (123 g/kg), arginine (80.6 g/kg) and aspartic acid (61.3 g/kg). While, Ullah *et al.*, (2016) and Ding *et al.*, (2018) they found that the content of amino acids was serine is (1.05 g/100 g), glutamic acid (3.50 g/100 g), glycine (0.95 g/100 g), alanine (1.05 g/100 g), lysine (0.97 g/100 g), and histidine (0.53 g/100 g). According to USDA (2018) the amino acids found in *S. hispanica* seeds, Glu is dominant (3.50 g/100 g dry seeds). Arg and Asp are also found in a high amount (2.14 and 1.69 g/100 g dry seeds, respectively), while others constitute less than 1 g/100 g dry seeds.

Table 7. Amino acids composition of proteins from black and white chia seeds genotypes (g/100g)

Amino acids (g/100g)	Black chia seeds	White chia seeds
Essential amino acids		
Arginine	6.97	7.6
Leucine	4.6	5.36
Lysine	3.37	4.08
Phenylalanine	3.6	3.8
Valine	3.44	3.52
Methionine	2.47	2.9
Threonine	2.44	2.77
Isoleucine	2.38	2.7

Histidine	1.93	2.27
Tryptophan	0.95	1.15
Total	27.55	36.15
Non-essential amino acids		
Glutamic	12.73	14.38
Aspartic	5.59	6.68
Alanine	4.31	4.26
Glycine	3.6	4.23
Serine	3.47	3.66
Proline	2.7	2.59
Tyrosine	2.51	2.84
Cystine	1.96	2.91
Total	36.87	41.55

Conclusions

Chia, *Salvia hispanica L.*, is a plant species used since ancient times for dietary and medical purposes. From the abovementioned results it can be concluded that the chia seeds contain a high protein, high fat content, dietary fiber, carbohydrates, minerals and antioxidants. Chia seeds contain the flavonoids, rosmarinic acid, quercetin, chlorogenic acid, and caffeic acid, which are proven to have anti-cancerogenic, anti-hypertensive, and neuron protective effects. Furthermore, chia seeds are a rich source of nutrients such as polyunsaturated ω -3 and ω -6 fatty acids that protect from inflammation, and antioxidant compounds that reduce the risk of chronic diseases. Moreover, the high amount of fiber decreases the risk of coronary heart disease, the risk for diabetes type 2, and several types of cancer. Finally, chia seeds are already used in the food and pharmaceutical industry. Also, chia seeds contain 18 out of 22 amino acids, of which contribute significantly to human health.

References

- Abdel-Aty, A.M.; Salama, W.H.; Fahmy, A.S. and Mohamed, S.A. (2019). Impact of germination on antioxidant capacity of garden cress: new calculation for determination of total antioxidant activity. *Scientia Horticulturae.*, 246: 155–160.
- Aguilera, Y.; Diaz, M.F.; Jimenez, T.; Benitez, V.; Herrera, T. and Cuadrado, C. (2013). Changes in non-nutritional factors and antioxidant activity during germination of non-conventional legumes. *J. Agri and Food Chem.*, 61: 8120-8125.
- Alcântara, M.A.; de Lima Brito Polari, I.; de Albuquerque Meireles, B.R.L.; de Lima, A.E.A.; da Silva Junior, J.C.; de Andrade Vieira, É.; dos Santos, N.A.; de Magalhães Cordeiro, A.M.T. (2019). Effect of the solvent composition on the profile of phenolic compounds extracted from chia seeds. *Food Chem.*, 275, 489–496.
- Álvarez-Jubete, L.; Wijngaard, H.; Arendt, E.K. and Gallagher, E. (2010). Polyphenol composition and in vitro antioxidant activity of amaranth, quinoa, buckwheat and wheat as affected by sprouting and baking. *Food Chem.*, 119: 770-778.
- AOAC. (2016). Official Methods of the Association of the Official Analytical Chemists. Maryland, USA.
- Ayerza, R. (2011). Seed yield components, oil content, and fatty acid composition of two cultivars of moringa (*Moringa oleifera* Lam.) growing in the Arid Chaco of Argentina. *Ind Crop Prod.*, 33:389-94.
- Beltrán-Orozco, M.C.; Martínez-Olgun, A. and Robles-Ramírez, M.C. (2020). Changes in the nutritional composition and antioxidant capacity of chia seeds (*Salvia hispanica L.*) during germination process. *Food Sci and Biotechnol.*, 29: 751–757.
- Berner, L.A.; Keast, D.R.; Bailey, R.L. and Dwyer, J.T. (2014). Fortified foods are major contributors to nutrient intakes in diets of US children and adolescents. *J. Academy of Nutri. and Dietetics.*, 114(7):1009-1022.
- Brand-Williams, W.; Cuvelier, M. E. and Berset, C. (1995). Use of a free radical method to evaluate antioxidant activity. *LWT-FoodTech.*, 28: 25–30.
- Carrillo, W.; Cardenas, M.; Carpio, C.; Morales, D.; Álvarez, M. and Silva M (2018). Content of nutrients component and fatty acids in chia seeds (*Salvia hispanica L.*) cultivated in Ecuador. *Asian J of Pharmaceutical and Clinical Res.*, 11(2):1-4.
- Chadare, F.J.; Idohou, R.; Nago, E.; Affonfere, M.; Agossadou, J.; Fassinou, T.K.; Kénou, C.; Honfo, S.; Azokpota, P. and Linnemann AR (2019). Conventional and food-to-food fortification: An appraisal of past practices and lessons learned. *Food Sci. and Nutri.* 7(9):2781-2795.
- Coelho, M.S. and Salas-Mellado, M. (2014). Chemical characterization of chia (*Salvia*

- hispanica* L.) for use in food products. *J. Food and Nutr. Res.*, 2(5):263-269.
- da Silva, B. P.; Anunciação, P. C.; Matyelka, J. C. da S.; Della Lucia, C.M.; Martino, H. S. D., and Pinheiro-Sant'Ana, H. M. (2017).** Chemical composition of Brazilian chia seeds grown in different places. *Food Chemistry.*, 221, 1709–1716.
- Das, A. (2018).** Advances in chia seed research. *Adv. Biotechnol. Microbiol.*, 5, 5–7.
- Di Marco, A.E.; Ixtaina, V.Y. and Tomás, M.C. (2020).** Inclusion complexes of high amylose corn starch with essential fatty acids from chia seed oil as potential delivery systems in food. *Food Hydrocolloids*, 108:106030.
- Dick, M.; Costa, T.M.; Gomaa, A.; et al. (2015).** Edible film production from chia seed mucilage: Effect of glycerol concentration on its physicochemical and mechanical properties. *Carbohydr Polym.*, 130: 198-205.
- Ding, Y.; Lin, H.W.; Lin, Y.L., et al. (2018).** Nutritional composition in the chia seed and its processing properties on restructured ham-like products. *J Food Drug Anal.*, 26(1): 124-34.
- Duranti, M. and P. Cerletti. (1979).** Amino acid composition of seed proteins of *Lupinus albus*. *J. Agric. Fd. Chem.* 27:977-978.
- Enes, B.N.; Moreira, Ld.P.D.; Toledo, R.C.L.; Moraes, É.A.; de Castro-Moreira, M.E.; Hermsdorff, H.H.M.; Noratto, G.; Mertens-Talcott, S.U.; Talcott, S. and Martino, H.S.D. (2020).** Effect of different fractions of chia (*Salvia hispanica* L.) on glucose metabolism, *in vivo* and *in vitro*. *J. of Functional Foods*, 71(4):104026.
- Gan, R.Y.; Wang, M.F.; Lui, W.Y.; Wu, K. and Corke, H. (2016).** Dynamic changes in phytochemical composition and antioxidant capacity in green and black mungbean (*Vigna radiata*) sprouts. *Inter. J. of Food Sci. and Technol.*, 51: 2090-2098.
- Ghafoor, K.; Ahmed, I.A.M.; Özcan, M.M.; Al-Juhaimi, F.Y.; Babiker, E.E. and Azmi, I.U. (2020).** An evaluation of bioactive compounds, fatty acid composition and oil quality of chia (*Salvia hispanica* L.) seed roasted at different temperatures. *Food Chem.*, 333(39):127531.
- Glass, R. L. (1971).** Alcoholysis, saponification and the preparation of fatty acid methyl esters. *Lipids*, 6(12): 919-925.
- Gómez-Favela, M.A.; Gutiérrez-Dorado, R.; Cuevas-Rodríguez, E.O.; Canizales-Román, V.A.; León-Sicairos, C.; Milán-Carrillo, J. and Reyes-Moreno, C. (2017).** Improvement of chia seeds with antioxidant activity, GABA, essential amino acids, and dietary fiber by controlled germination bioprocess. *Plant Foods Hum. Nutr.*, 72: 345-352.
- Grancieri, M.; Martino, H.S.D.; Gonzalez de Mejia, E. (2019).** Chia Seed (*Salvia hispanica* L.) as a Source of Proteins and Bioactive Peptides with Health Benefits: A Review. *Compr. Rev. Food Sci., Food Saf.*, 18, 480–499.
- Hakkinen, S. H.; Karenlampi, S. O.; Heinonen, I. M.; Mykkanen, H. M. and Torronen, A. R. (1998).** HPLC method for screening of flavonoids and phenolic acids in berries. *J. Sci. Food Agric.*, 77: 543-551.
- Hasler, C.M. and Brown, A.C. (2009).** Position of the American Dietetic Association: Functional foods. *J Am Diet Assoc.*, 109(4): 735-46.
- Ixtaina, V.Y.; Vega, A.; Nolasco, S.M.; Tomás, M.C.; Gimeno, M.; Bárzana, E. and Tecante, A. (2010).** Supercritical carbon dioxide extraction of oil from Mexican chia seed (*Salvia hispanica* L.): Characterization and process optimization. *J. Supercrit. Fluids*, 55, 192–199.
- Jin, F.; Nieman, D.C.; Sha, W.; Xie, G.; Qiu, Y. and Jia, W. (2012).** Supplementation of milled chia seeds increases plasma ALA and EPA in postmenopausal women. *Plant Foods Hum. Nutr.*, 67, 105–110.
- Kim, D.K.; Jeong, S.C.; Gorinstein, S. and Chon, S.U. (2012).** Total polyphenols, antioxidant and antiproliferative activities of different extracts in mungbean seeds and sprouts. *Plant Foods Hum. Nutr.*, 67: 71-75
- Knez-Hrnčić, M.; Cör, D. and Knez, Ž. (2018).** Subcritical extraction of oil from black and white chia seeds with n-propane and comparison with conventional techniques. *J. Supercrit. Fluids*, 140, 182–187.
- Knez-Hrnčić, Ivanovski, M.; Cör, D. and Knez, Ž., (2019).** Chia seeds (*Salvia hispanica* L.): An overview—Phytochemical profile, isolation methods, and application. *Molecules*, 25, 11.
- Kulczynski, B.; Kobus-Cisowska, J.; Taczanowski, M.; Kmiecik, D. and Gramza-Michalowska A. (2019).** The chemical composition and nutritional value of chia seeds-current state of knowledge. *Nutr.*, 11: 1242.
- Kuznetcova, D.V.; Linder, M.; Jeandel, C.; Paris, C.; Desor, F.; Baranenko, D.A.; Nadtochii, L.A.; Arab-Tehrany, E.; Yen, F.T. (2020).** Nanoliposomes and nanoemulsions based on chia seed lipids: Preparation and characterization. *Int. J. Mol. Sci.*, 21, 9079.
- Marineli, R. da S.; Lenquiste, S. A.; Moraes, É. A.; Maróstica, M. R. (2015).** Antioxidant potential of dietary chia seed and oil (*Salvia hispanica* L.)

- in diet-induced obese rats. *Food Research International*, 76: 666–674.
- Martínez-Cruz, O. and Paredes-López, O. (2014).** Phytochemical profile and nutraceutical potential of chia seeds (*Salvia hispanica L.*) by ultra-high performance liquid chromatography. *J. Chromatogr., A*, 1346: 43-48.
- Mattila, P.; Astola, J. and Kumpulainen, J. (2000).** Determination of flavonoids in plant material by HPLC with diode-array and electro-array detections. *J. Agric. Food Chem.*, 48: 5834-5841.
- Mohd Ali, N.; Yeap, S.K.; Ho, W.Y.; Beh, B.K.; Tan, S.W.; Tan, S.G. (2012).** The promising future of chia, *Salvia hispanica L.* *J. Biomed. Biotechnol.*, 1–9.
- Motyka, S.; Koc, K.; Ekiert, H.; Blicharska, E.; Czarnek, K.; Szopa, A. (2022).** The Current State of Knowledge on *Salvia hispanica* and *Salvia hispanica* semen (Chia Seeds). *Molecules*, 27, 1207.
- Oliveira-Alves, S.C.; Vendramini-Costa, B.D.; BaúBetimCazarin, C.; Maróstica, M.R.; Ferreira, J.P.B.; Silva, A.B.; Prado, M.A. and Bronze, M.R. (2017).** Characterization of phenolic compounds in chia (*Salviahispanica L.*) seeds, fiber flour and oil. *Food Chem.*, 232: 295–305.
- Olivos-Lugo, B.L.; Valdivia-Lopez, M.A. and Tecante, A. (2010).** Thermal and physicochemical properties and nutritional value of the protein fraction of Mexican chia seed (*Salvia hispanica L.*). *Food Sci Technol Int.*, 16: 89-96.
- Otondi, E.A.; Nduko, J.M. and Omwamba, M. (2020).** Physico-chemical properties of extruded cassava-chia seed instant flour. *J. Agri. and Food Res.*, 2:100058.
- Pellegrini, M.; Lucas-Gonzalez, R.; Sayaas-Barberá, E.; Fernández-López, J.; Pérez-Álvarez, J. A., 768 and Viuda-Martos, M. (2018).** Bioaccessibility of Phenolic Compounds and Antioxidant Capacity of Chia (*Salvia hispanica L.*) Seeds. *Plant Foods for Human Nutr.*, 73, 47-53.
- Poudyal, H.; Panchal, S.K.; Waanders, J.; Ward, L. and Brown, L. (2012).** Lipid redistribution by α -linolenic acid-rich chia seed inhibits stearoyl-CoA desaturase-1 and induces cardiac and hepatic protection in diet-induced obese rats. *J. Nutr. Biochem.* 23: 153–162.
- Rahman, M.J.; de Camargo, A.C. and Shahidi, F. (2017).** Phenolic and polyphenolic profiles of chia seeds and their in vitro biological activities. *J. Funct. Foods*, 35, 622–634.
- Rajaram, S. (2014).** Health benefits of plant-derived α -linolenic acid. *The Ameri. J. Clini. Nutr.*, 100:443S-448S.
- Re, R.; Pellegrini, N.; Proteggente, A.; Pannala, A.; Yang, M. and Rice-Evans, C. (1999).** Antioxidant activity applying an improved ABTS radical cation decolorization assay. *Free Radical Biol. Med.*, 26: 1231-1237.
- Reyes-Caudillo, E.; Tecante, A. and Valdivia-Lopez, M.A. (2008).** Dietary fibre content and antioxidant activity of phenolic compounds present in Mexican chia (*Salvia hispanica L.*) seeds. *Food Chem.*, 107: 656-63.
- Ritthibut, N.; Oh, S. and Lim, S. (2021).** Enhancement of bioactivity of rice bran by solid-state fermentation with *Aspergillus* strains. *LWT Food Sci Technol.*, 135:110273.
- Shi, H.L.; Nam, P.K. and Ma, Y.F. (2010).** Comprehensive profiling of isoflavones, phytosterols, tocopherols, minerals, crude protein, lipid, and sugar during soybean (*Glycine max*) germination. *J. Agri. and Food Chem.*, 58: 4970-4976.
- Singleton, V. L. and Rossi, J. A. (1965).** Colorimetry of total phenolics with phosphomolybdic-phosphotungstic acid reagents. *Am. J. Enol. Vitic.*, 16: 144-158.
- Suri, S.; Passi, S.J. and Goyat, J. (2016).** Chia seed (*Salvia hispanica L.*). A new age functional food. In: 4th International Conference on Recent Innovations in Science Engineering and Management, 286-299.
- Tamime A.Y.; Muir, D.D.; Barclay, M.N.I.; Khas-Kheli, M. and McNulty, D.(1997).** Laboratory-made Kishk from wheat, oat and barley: 2. Compositional quality and sensory properties. *Food Res Int.*, 30:319–326
- Timilsena, Y.P.; Adhikari, R.; Barrow, C.J. and Adhikari, B. (2017).** Digestion behaviour of chia seed oil encapsulated in chia seed protein-gum complex coacervates. *Food Hydrocoll.*, 66, 71–81.
- Ullah, R.; Nadeem, M.; Khalique, A.; Imran, M.; Mehmood, S.; Javid, A. and Hussain, J. (2016).** Nutritional and therapeutic perspectives of Chia (*Salvia hispanica L.*): A review. *J. Food Sci. Technol.*, 53, 1750–1758.
- USDA. (2018).** USDA National Nutrient Database for Standard Reference Full Report, Seeds, chia seeds.
- Zhishen, J.; Mengcheng, T. and Jianming, W. (1999).** The determination of flavonoid contents in mulberry and their scavenging effects on superoxides radicals. *Food Chem.*, 64: 555-559.

تقييم المكونات الحيوية النشطة والخواص المضادة للأكسدة لنوعين من بذور الشيا (زنجبيل كجسبنيك).
 **إشراق بدوي كامل- *فرحات فوده على فوده- *إبراهيم محمد عبد العليم- *مني عبد المنعم محمد علي
 * قسم الكيمياء الحيوية الزراعية - كلية الزراعة - جامعة بنها-مصر
 ** معهد بحوث تكنولوجيا الاغذية- مركز البحوث الزراعية - الجيزة-مصر

الهدف من هذا البحث هو تقييم بذور الشيا السوداء (*Salvia hispanica L.*) كمصدر للمكونات الحيوية النشطة مقارنة مع بذور الشيا البيضاء. وأيضاً تقدير المكونات الكيميائية الحيوية وكذلك محتواها من المعادن ومحتواها من المواد المضادة للأكسدة، الفينولات والفلافونيدات وتفردها والتعرف عليها باستخدام جهاز التحليل الكروماتوجرافي العالي (HPLC) والأحماض الدهنية المشبعة وغير المشبعة والأحماض الأمينية ومعرفة الأساسية منها والغير أساسية.

أظهرت النتائج اختلافات طفيفة في التركيب الكيميائي بين بذور الشيا الخام (السوداء والبيضاء) وارتفاع القيمة الغذائية لها متمثلة في محتواها من البروتين (25.29 - 24.30%) ، ألياف (32.94 - 31.75%) ، دهون (32.35 - 30.54%) ومحتوى رماد. (6.05-6.93%) على التوالي. بالنسبة لبذور الشيا النابتة ، أظهرت البيانات أن محتوى البروتين في بذور الشيا البيضاء زاد معنوياً 26.94% مقارنة ببذور الشيا البيضاء الخام 24.30%. بينما وجد أن محتوى الدهون والألياف انخفض بشكل ملحوظ نتيجة لعملية الإنبات. أظهرت بذور الشيا الممتخمة محتوى بروتيني أعلى بنسبة 28.10 و 27.55% لبذور الشيا البيضاء والسوداء مقارنة ببذور الشيا الخام. تم تعريف المركبات الفينولية الكلية بواسطة جهاز التحليل الكروماتوجرافي العالي (HPLC) ومنها حمض الكافيك، الروزمارينيك وحمض إيلاجيك بين نوعي بذور الشيا. زاد محتوى الفينولات الكلية بشكل معنوي في البذور السوداء النابتة (263.51 ± 28.44 مللجم / 100 جم جاليك اسيد) مقارنة بالبذور الخام (166.76 ± 4.13 مللجم / 100 جم جاليك اسيد) بينما أظهرت البذور الممتخمة زيادة معنوية في محتوى الفينولات الكلية لكل من نوعي بذور الشيا حيث كانت القيم 216.25 ± 4.75 و 200.74 ± 3.64 مللجم / 100 جم جاليك اسيد عند مقارنتها بالعينة الخام. تحتوي كلا من نوعي بذور الشيا على أحماض أمينية أساسية بنسب متقاربة. كان النشاط المضاد للأكسدة الأعلى نسبياً في بذور الشيا السوداء الخام مقارنة ببذور الشيا البيضاء الخام. إلى جانب ذلك ، يحتوي زيت الشيا على نسبة عالية من حمض اللينولينيك (ω 3) بقيمة 59.25 - 58.54% من إجمالي محتوى الأحماض الدهنية في زيت بذور الشيا السوداء والبيضاء على التوالي. محتوى حمض اللينولينيك (ω 6) كان من 20.50-20.17%. وقد تبين من هذه الدراسة احتواء بذور الشيا علي العناصر الغذائية بنسب متوازنة وكذا تعد كمصدر غني بالمعادن، مضادات الأكسدة، الأحماض الدهنية غير المشبعة، الأحماض الأمينية الأساسية والتي لها دور في الحماية من الالتهابات وتقليل مستوى الكوليسترول وتقليل خطر الإصابة بالنوبات القلبية والسرطان وتوفير الحماية ضد مرض السكري من النوع الثاني.