

Effect of Different Dietary Copper Forms and Levels on Carcass Characteristics and Meat Quality Traits of Broilers Chickens

Ibrahim, I. A.¹; G. M. EL-Gendi¹; A. A. Nihad²; H. M. Okasha¹ and M. M. El-Attrouny¹

¹Animal Production Dept., Fac. of Agric., Benha Univ., Egypt.

²College of Agriculture, Al-Qasim green University. Iraq.

Abstract

This study was carried out to evaluate the effects of dietary copper supplementation at a levels of 75, 100 and 125 ppm, from various forms, inorganic copper sulfate, (CuSo₄) and copper nanoparticles (Cu-NP) supplementation on the live body weight, carcass characteristic and meat quality traits of broiler chickens. A total of 420 one-day-old Ros 308 chicks were randomly assigned to seven treatments. (n= 60 birds per each). The 1st group was fed a basal diet without any supplementation and considered as control group, the 2nd, 3rd and 4th groups were fed basal diet supplemented with 75, 100 and 125 ppm / kg diet of CuSo₄, the 5th, 6th and 7th groups were fed basal diet supplemented with Cu-NPs at a level 75, 100 and 125 ppm / kg diet. Results showed that birds fed basal diet supplemented with inorganic or Cu-NPs had significantly better live body weight, carcass characteristics and meat quality while fat percentage was reduced compared with control group. However, the groups supplemented with Cu-NP showed significant improvement on carcass and meat quality traits compared with the other groups. Supplementation of dietary Cu from different forms significantly decreased meat content fat, PH, TVN and TBA. On the other hand, supplementation of dietary Cu from different forms significantly increased the levels meat sensory evaluation (color and Elasticity) compared with the control group. Dietary cu supplementation led to the total monounsaturated fatty acid (TMUFA) and total polyunsaturated fatty acid (TPUFA) content of muscles increased and total saturated fatty acids decreased compared with the control group. In conclusion, Cu supplementation with 125 ppm/ kg diet of Cu-NPs improved the live body weight, dressing percentage, and meat chemical examination of broiler chickens. Broilers fed Cu-NP had better growth and meat chemical examination than those fed CuSo₄.

Keywords: copper, carcass characteristics, meat quality, nanoparticles, fatty acid, broilers

Introduction

A major issue in the meat industry is a decrease in the acceptability and nutritional quality of meat, as well as changes in flavor caused by the lipid oxidation process. As a result of changes and improvements in peoples eating habits, there is a growing interest in natural health and functional foods, including the development of new convenience meat products with natural flavor and taste (Choi *et al.*, 2010).

Copper (Cu) a crucial microelement required for proper physiological and biochemical processes, is frequently added to poultry diets at high concentrations at levels often exceed the birds requirements (Boa and Chocht, 2009; Świątkiewicz *et al.*, 2014), copper as a possible alternative to an antibiotic growth promoters. It enhances animal performance, but an excess of Cu in the diet can also have adverse effects, including iron and calcium deficiency due to antagonism between those elements; this may cause a reduction in viability (Miroshnikova *et al.*, 2015) and increased toxicity (Cao *et al.*, 2016). The digestibility of Cu salts is very low, and approximately 80% of Cu is excreted in the feces (McDowell, 1992). Copper nanoparticles (Cu-NP) are relatively more bioavailable due to their small size and high surface to volume ratio. Although, there is limited research on the use of Cu-NP in the poultry industry. Some researchers have

pointed out the importance of Cu-NP as an effective alternative of chemical, anti-bacterial agents, and growth promoters (Mroczek *et al.*, 2015; Joshua *et al.*, 2016; sharif *et al.*, 2021). It has been documented that Cu-NP has beneficial effects on animal performance and could be used to replace copper sulfate (CuSO₄) (Wang *et al.*, 2011; Mroczek *et al.*, 2016; Muralisankar *et al.*, 2016; El-Basuini *et al.*, 2016). However, far less is known about the mechanism of action of Cu-NP in improving chicken performance, particularly regarding nutrient digestion and metabolism. Increasing Cu levels in the diet could reduce cholesterol and triglyceride levels in the blood of chickens (Rahman *et al.*, 2001; Skrivanova *et al.*, 2000; Ibrahim *et al.*, 2022). Therefore, this study aimed to investigate the effects of feeding different copper forms (CuSo₄ and Cu-NPs) on carcass characteristics, meat quality traits (chemical composition - sensory evaluation, and fatty acids profile) of broiler chickens.

Materials and Methods

Experimental Location and Ethics approval

The Poultry Research Farm, Faculty of Agriculture at Benha University, Egypt served as the study's location. The Institutional Animal Care and Use Committee (IACUC) at Benha University

authorized all experimental protocols. During the period from 21 May to 26 June 2021.

Experimental design and dietary treatments:

A completely randomized design involving a 2×4 factorial arrangement of treatments was used in this study. Four supplemental levels including 0, 75, 100 and 125 mg/kg copper from the two copper forms (copper sulfate and copper nanoparticles). Seven diet were formulated. Diet one without Cu supplementation and consider as a control diet. Diet

2-4 were basal diets supplemented with 75, 100, 125 mg/kg diet copper sulfate, respectively. While, Diet 5-7 were basal diet supplemented with 75, 100, 125 mg/kg diet Cu-NPs. Table 1 shows the basal and grower diets were formulated according to NRC (1994). The trial lasted 5 weeks. Mash diet and fresh water were provided *ad-libitum*. All chicks were reared under similar managerial and hygienic conditions.

Table 1. Feed ingredients and nutritional value of the basal starter and grower diet.

Ingredients (%)	Starter (1-21) days	Gower (22-35) days
Yellow Corn	58.0	63.0
Soybean (44% CP) ^a	28.2	24.9
Gluten meal (60% CP) ^b	7.00	6.00
Dicalcium phosphate	1.80	1.20
Limestone	1.30	1.30
Soybean oil	2.00	2.00
NaCl	0.30	0.30
L-Lysine hydrochloride	0.20	0.20
DL-Methionine	0.20	0.10
Mineral Premix*	0.50	0.50
Vitamin premix**	0.50	0.50
Total	100	100
Nutrient analysis		
ME (kcal/kg) ^d	3012	3069
Crude protein (%)	21.99	20.28
Calcium (%)	0.98	0.84
Total phosphor (%)	0.72	0.59
Methionine (%)	0.59	0.47
Methionine + Cystine (%)	0.95	0.80
Lysine (%)	1.14	1.06

^{a,b}Crude protein. ^cProvided the following per kilogram of diet: 13,000 IU of vitamin A; 1,300 IU of vitamin D; 65 IU of vitamin E; 3.4 mg of menadione; 37 mg of pantothenic acid; 6.6 mg of riboflavin; 3.7 mg of folic acid; 39 mg of niacin; 1.0 mg of thiamine; 4.3 mg of vitamin B6; 0.23 mg biotin; 0.075 mg of vitamin B12: 43 mg of choline chloride. 170 mg of zinc; 140 mg of iron; 34 mg of manganese; 16 mg of copper; 0.29 mg of iodine; 0.29 mg of selenium. ^dMetabolizable energy.

A total of 420 one-day old male (Ross 308) chicks were individually assigned randomly to seven groups, each with three replicates of 20 birds each based on a completely randomized design. Thermo-neutral ambient temperature was maintained in accordance to standard brooding practices and adapted to the birds rearing stages. Light regime was regulated as follows 24 h light (1st-5th day) and 23h light and 1 h dark (6th-35th day) used in the present experiment. Floor brooder with gas heaters were used to provide chicks with heat needed for brooding. The brooding temperature was 32, 30, 29 and 23-26 °C at 1-7, 8-14, 15-20 and 21-35 days of age, respectively (gradually decreased).

Slaughtering and carcass measurements

At the end of the experiment (35 days of age), 5 birds from each treatment were individually weighed, and slaughtered by severing the jugular vein with a sharp

knife near the first neck vertebra. Chickens were fasted for a period of 8-hours before slaughter,

however, they had unlimited access to water. After slaughter and bleeding, the carcasses were de-feathered and eviscerated and intestine, gizzard, lungs, spleen, liver, heart and all internal organs were removed. The carcass and giblets (empty gizzard liver and heart) were separately weighed and expressed as a percentage of live body weight. The proportional weights to live weight of giblets, carcass and total edible parts were calculated as follows: giblets weight (%) = (GW/LW) ×100, edible parts (%) = ((EW+GW)/LW) ×100, whereas: LW = live weigh, EW= eviscerated weigh and GW= giblets weight. the breast and thigh meat was sampled. Part of the meat was immediately used for the determination of pH, moisture, protein, fat, and ash, while the remainder was frozen at -20 °C for subsequent analysis (total lipids and fatty acid profile).

Meat quality traits

Chemical composition

The examined samples of chicken fillets and sheish were analyzed for determination of their contents of moisture, protein, fat and ash by using the standard method recommended by Horwitz (2000) "AOAC". Keeping quality tests by determination of pH (Pearson 2006), total volatile nitrogen (TVN) was recommended by Food and Agriculture Organization FAO (1980) as follows: $TVN/100g = 26.88 \times (2-T1)$. Where, T1 = volume of NaOH consumed in the titration. Thiobarbituric acid number (TBA) by Pikul *et al.*, (1989) was applied as follow: $TBA \text{ value} = R \times 7.8$ (mg malonaldehyde /Kg). Where, R = Reading of sample against blank.

Sensory evaluation

The examined samples of chicken meat were analyzed for the quantification of the final sensory profile according to procedures of the World's Poultry Science Association by Mead (1987). Five trained panelists applied the proposed organoleptical method of raw chicken meat analysis. The different attributes were quantified on a rating scale from 1 to 3. The sensorial analyzed attributes were visual look (skin and meat color), meat consistency and elasticity and the odor.

Fatty Acids Profile of Meat

One hundred grams of the meat sample were placed in a 500 ml closed stopper flask then, 300 ml of n-hexane was added, and the flask was shaken for 30 min. using horizontal shaker and left for 24 hours at room temperature. The homogenated mixture was filtered and the residue was re-extracted as mentioned above. The combined filtrates were evaporated under reduced pressure, according to AOAC (2005). Fatty acids were determined in meat by Gas Chromatography technique (GC) according to Aura *et al.* (1995). The fats under study were saponified with ethanolic potassium hydroxide (40%, w/v) for 24 hours at room temperature according to the method of AOCS (1993). The aqueous layer (containing potassium salt of fatty acids and free from unsaponifiable matter) was acidified with HCL (0.5N), then it was extracted three time with petroleum ether. The petroleum ether extract was washed several times with distilled water and dried over anhydrous sodium sulphate. The obtained fatty acids were converted to methyl esters as follows: The extracted fatty acids were dissolved in anhydrous diethyl ether (0.5-1.0ml) and methylated by drop wise addition of diazomethane solution (Vogel, 1975) until the yellow color persisted. The mixture was then left at room temperature for 15 minutes and the solvent was evaporated on a water bath maintained at 60C. Finally, the methyl ester of fatty acids was dissolved in chloroform and aliquots of this solution were subjected to analysis by GC. The fatty acids methyl esters were analyzed by Hewlett Packard gas chromatography (5890 series) equipped with flame ionization detector. The chromatograph

was fitted with FFAP (2.5m × 0.30µm film thickness and 0.32mm diameter).

Statistical analysis

Two-way ANOVA was used to examine the data, using the General Linear Models (GLM) method of (SAS, 2004). A factorial design (2×3) was used to assess the interaction between the main effects elements (dietary copper forms and amount). Duncan's multiple range test was applied to evaluate differences among treatments, where significant differences (Duncan 1955). According to the following liner model:

$$X_{ijk} = \mu + S_i + L_j + (SL)_{ij} + e_{ijk}$$

Whereas: X_{ijk} = the observation of traits for ijk^{th} birds; μ = the overall mean.; S_i = the effect of the i^{th} copper forms; L_j = the effect of the j^{th} copper levels.; $(SL)_{ij}$ = the fixed effect of the interaction between the i^{th} sources and the j^{th} levels.; e_{ij} = random error assumed to be independently and randomly distributed.

Results and discussion

Live body weight and carcass characteristics.

Results of the proportional weights of carcass, giblets and total edible parts for broiler chickens as affected by copper forms, copper levels and the interaction between them are presented in Table (2). All copper forms and levels supplementation created no significant ($P < 0.05$) difference found in body weight (BW) at 5 wks of age on absolute wights of carcass, heart, and total edible parts. Which increased in chicks However, the previous treatments were superior for the groups fed diet supplemented with Cu -NPs compared with fed diets supplemented with copper sulfate.

Copper levels showed a significantly, ($P < 0.05$) increased body weight and carcass weight percentage at 125 mg/kg diet. In addition, broiler chickens fed diet supplemented with 100 mg/kg diet from either Cu -NPs significantly, ($P < 0.05$) increased absolute weight edible parts when compared with different levels applied and control group. Similar to our results, it was shown that supplementation with 100 mg/kg of Cu-NP significantly enhanced growth performance in piglets (Gonzales *et al.*, 2009; Wang *et al.*, 2012). Furthermore, similar effects of Cu-NP on the final BW in fish were reported (El-Basuini *et al.*, 2016). Our results are in agreement with a study on broilers (Scote *et al.*, 2016), demonstrating that the improvement in broilers growth could be attributed to better energy digestibility in animals treated with Cu-NP than with $CuSO_4$.

Previously, we demonstrated that 125 mg/kg of Cu-NP had significant effects on the broiler dressing percentage and carcass content (Mroczek-Sosnowska *et al.*, 2015). The same authors reported that Cu supplementation reduced the heart weight. However, our results showed that the relative heart weight was

not affected. Similar to our results, it was reported that the relative heart weight at 42 days age was not affected in birds treated with CuSO₄ (Shahzad *et al.*, 2012). The relative liver weight was affected by both forms of Cu and was significantly lower at higher concentrations of Cu in relation to the control group. The liver results are consistent with the results of Shahzad *et al.*, 2012; Skrivan *et al.*, 2000), while in other studies (Mroczek- Sosnowska *et al.*, 2015; Payvastegan *et al.*, 2013; Upadhaya *et al.*, 2016; Wang *et al.*, 2011), the liver weight was not affected. It was reported that a high concentration of Cu-NP negatively affected the histology of the liver, kidneys and spleen, but not the heart and lungs in rats, and these changes were well supported by organ weight changes (Lee *et al.*, 2016). Furthermore, it was demonstrated that the greatest accumulation of Cu was observed in the liver and spleen (Mroczek-Sosnowska *et al.*, 2014).

Chemical examination of meat

Results presented in Table (3) showed that highly significant ($P < 0.05$) effect was found on moisture, protein, fat and ash, respectively due to copper forms. Concerning to the effect of the copper levels on meat chemical composition it is clearly found that there was highly significant ($P < 0.001$) effect on protein, fat and ash of meat due to copper levels. Birds fed diets supplemented with CuSO₄ and Cu-NPs at different levels had significantly ($P < 0.001$) higher values of Moisture, protein, and ash % than the control group, with the higher fat % value observed in control group compared to the other treated groups (Table 3).

Meat sensory evaluation

PH, TVN and TBA increased significantly in meat samples produced from chickens fed diet supplemented Cu-sulfate compared with those produced from Cu-NPs. PH, TVN and TBA decreased significantly with increasing Cu levels, reaching its maximum decreasing at 125 mg / kg diet when compared with control group, which showed the higher values of the previously mentioned traits (table, 4)

Results in Table 4 and 5 showed significant ($P < 0.05$) effect on values of aspect, odor, color, elasticity and overall score values in sensory examination of broiler meat the higher values of color (2.77), elasticity (2.44) and overall score (9.88) were found significant Cu-NPs than Cu-So₄ (Bianchi *et al.*, 2006 and Boni *et al.*, 2010). Highly significant ($P < 0.001$) effects on odor and overall score values in meat sensory examination were observed in the groups fed of Cu at a level of 100 and 125 mg/kg diet (table, 5). These results agree with those reported by Dadgar *et al.* (2010) who found that birds subjected to conditions of hypoxia before slaughter resulted in darker-colored breast meat .

The sensory evaluation results showed an overall improvement in order, color and overall. It could be ascribed to the antioxidant properties of different copper forms that inhibited lipid oxidation of meat, which is known to be the main reason behind meat quality spoiling resulting in rancidity and formation of unacceptable odors and flavors (Amaral *et al.*, 2018). Generally, there was an improvement in meat elasticity expressed in response to copper forms.

Changes in Fatty Acid Composition

Fatty acid compositions of broiler thigh and breast muscle produced from dietary different copper forms and levels supplementation are presented in Table 6. There were some differences in the percentages of palmitoleic acid (C16:0) and palmitoleic acid (C16:1) among all treatments. Also, significant differences were observed in the percentages of myristic acid (C14:0), stearic acid (C18:0), Oleic (C18:1), Linoleic (C18:2), Linoleic (C18:3) and arachidonic acid (C20:4) among all treatments due to all factors studied. Overall percentages of oleic (C18:1), linoleic (C18:2), and linolenic acid (C18:3) significantly increased ($P < 0.05$). (Choi *et al.* (2010); López-Ferrer *et al.*, 2001). Furthermore, there was a numeric decrease in the percentages of palmitic acid (C16:0) with incremental dietary levels of Cu-NP (100 and 125 mg/ kg diet) and this result was significant ($P < 0.05$) in the treatment with 125 Cu-NP compared with other treatment .The highest concentrations of unsaturated FAs (both MUFAs and PUFA) were present in the diet supplemented with Cu-NP a level of 100 and 125 mg. In contrast, the highest concentration of SFA was present in the diet supplemented without copper. There was important effect of Cu supplementation on the fatty acid profile of lipids of breast and thigh muscles, except minor changes in contents of some fatty acids. Thus, the reduction of total muscle lipids and cholesterol content was the main effect of Cu supplementation in chickens (Kim *et al.*, 2005; Guo *et al.*, 2006).

Conclusions

The results can be summarized that the supplementation of Cu-NP and CuSo₄ to broiler chicken diets improved body weight, carcass characteristics and meat quality traits. Birds fed diet with Cu-NP copper improved productive performance than other groups. Also, increasing the levels of Cu led to decreased total saturated fatty acid, and increased total monounsaturated fatty acid and total polyunsaturated fatty acid.

Table 2. Relative weights of carcass components as affected by the different levels of dietary copper supplementation (% of live body weight).

Items	Levels (mg/kg)	Live body weight	Carcass percentage	Gizzard (%)	Liver (%)	Heart (%)	Giblets (%)	Edible part (%)
Sources	Cu - NP	1958±.55	75.9±1.5	1.98±0.09	2.23±0.08	0.55±0.03	4.77±0.13	80.7±1.4
	Cu sulfat	1938±55	74.8±1.5	2.14±0.09	2.27±0.08	0.54±0.03	4.96±0.13	79.7±1.4
Copper level	0	1875±63b	72.5±1.6 c	1.85±0.11b	2.20±0.09	0.48±0.06	4.53±0.15	77.3±1.5c
	75	1936±.63a	72.5±1.6 c	2.22±0.11a	2.13±.0.09	0.61±0.06	4.98±0.15	77.3±1.5c
	100	1877±63a	75.0±1.6 b	1.97±0.11a	2.34±0.09	0.53±0.06	4.86±0.15	83.5±1.5a
Control	125	2032±63a	78.6±1.6 a	1.97±0.11a	2.29±0.09	0.50±0.06	4.77±0.15	79.9±1.5b
	0	1847±.0. 94	72.5±1.8	1.85±0.16b	2.20±0.13	0.48±0.03b	4.53±.0.2 2	77.3±0.57c
Cu -NP	75	1918±0. 94	72.9±1.8	2.20±0.16a b	2.09±0.13	0.54±0.03a b	4.92±0.22	77.8±0.84ab
	100	1898±0.94	75.7±1.8	1.79±0.16a	2.31±0.13	0.53±0.03a b	4.63±0.22	80.8±0.84a
CuSo4	125	2060±.0.9 4	75.2±1.8	1.95±0.16a	2.29±.0.1 3	0.50±0.03a b	4.76±.0.2 2	80.3±0.84ab
	75	1954±0.94	72.0±1.8	2.25±0.16a b	2.16±0.13	0.60±0.03a	5.03±0.22	76.8±0.84c
	100	1856±0.94	77.7±1.8	2.16±0.16a b	2.38±0.13	0.53±0.03a b	5.09±0.22	78.9±0.84ab c
	125	2004±.0.9 4	74.8±1.8	1.99±0.16a	2.28±.0.1 3	0.50±0.03a b	4.78±.0.2 2	79.6±0.84ab c

^{a,b,c} Means with different superscript in the same column are significantly different at (P<0.05).

CuSo4 = Copper sulfate; Cu-NP = Copper nanoparticles.

Table (3): Least – square means and standard error for meet chemical examination (moisture, protein, fat and ash) as affected by the studied factors

Items	Levels (mg/kg)	Moisture (%)	Protein (%)	Fat (%)	Ash (%)
Sources	Cu -NP	74.5±0.11a	20.2±0.18a	2.04±0.08b	2.03±0.05a
	Cu sulfate	74.3±0.11b	19.4±0.18b	2.42±0.08a	1.63±0.05b
Copper level	0	74.2±0.20a	18.90±0.35b	2.86±0.17a	1.43±0.14b
	75	74.5±0.14a	19.51±0.25ab	2.40±0.12b	1.71±0.10ab
	100	74.4±0.14a	19.86±0.25a	2.21±0.12b	1.83±0.10a
Control	125	74.3±0.14a	20.25±0.25a	2.08±0.12b	1.95±0.10a
	0	74.2±0.18a	18.9±0.29c	2.86±0.14a	1.43±0.09d

	75	74.6±0.18cd	19.8±0.29abc	2.23±0.14bcd	1.86±0.09bc
Cu -NP	100	74.2±0.18ab	20.20±0.29ab	2.03±0.0.14cd	2.03±0.09ab
	125	74.1±0.18a	20.7±0.29a	1.86±0.0.14d	2.20±0.09a
	75	74.4±0.18de	19.2±0.29c	2.56±0.0.14ab	1.56±0.09d
CuSo4	100	74.6±0.18cd	19.5±0.29c	2.40±0.0.14bc	1.63±0.09cd
	125	74.6±0.18bc	19.7±0.29bc	2.3±0.14bcd	1.70±0.09cd

^{a-d} Means with different superscript in the same column are significantly different at (P<0.05).

CuSo4 = Copper sulfate; Cu-NP = Copper nanoparticles.

Table 4 Least – square means and standard error for meet chemical examination (Ph, TVN and TBA) as affected by the studied factors

	Levels (mg/kg)	pH	TVN (mg/Kg)	TBA (mg/Kg)
	Cu -NPs	5.56±0.009b	3.13±0.16b	0.11±0.005b
Sources	Cu sulfate	5.63±0.009a	3.91±0.16a	0.15±0.005a
	0	5.67±0.02a	4.66±0.31a	0.19±0.18a
Copper level	75	5.62±0.01ab	3.91±0.22b	0.14±0.13b
	100	5.60±0.01b	3.51±0.22bc	0.13±0.13a
	125	5.57±0.01b	3.13±0.22c	0.11±0.13a
	Control	0	5.67±0.01a	4.66±0.21a
	75	5.60±0.01d	3.66±0.21bc	0.13±0.01bc
Cu -NP	100	5.56±0.01e	3.16±0.21cd	0.11±0.01bc
	125	5.53±0.01e	2.56±0.21d	0.09±0.01c
	75	5.65±0.01ab	4.16±0.21ab	0.16±0.01ab
CuSo4	100	5.63±0.01bc	3.86±0.21bc	0.15±0.01ab
	125	5.61±0.01cd	3.70±0.21bc	0.14±0.01abc

^{a-d} Means with different superscript in the same column are significantly different at (P<0.05).

CuSo4 = Copper sulfate; Cu-NP = Copper nanoparticles.

Table 5. Least - square means and standard error for meet sensory evaluation (aspect, older, color and elasticity) as affected by the studied factors.

Items	Levels (mg/kg)	Aspect (3)	Oder (3)	Color (3)	Elasticity (3)	Overall (12)
Sources	Cu -NPs	2.77±0.16	2.88±0.14	2.77±0.16a	2.44±0.20a	9.88±0.36a
	Cu sulfate	2.55±0.16	2.66±0.14	2.44±0.16b	2.22±0.20b	9.88±0.36b
Copper level	0	2.33±0.29	1.66±0.26b	1.66±0.28b	2.00±0.38	7.66±0.67b
	75	2.50±0.20	2.66±0.18a	2.33±0.20ab	2.16±0.27	9.66±0.47a
	100	2.66±0.20	2.83±0.18a	2.66±0.20a	2.33±0.27	10.6±0.47a
	125	2.83±0.20	2.83±0.18a	2.83±0.20a	2.50±0.27	11.0±0.47a
Control	0	2.33±0.19	1.66±0.02c	1.66±0.02b	2.00±0.02	7.66±0.57c
	75	2.66±0.19	2.66±0.02b	2.33±0.02ab	2.33±0.02	10.00±0.57ab
Cu-NP	100	2.66±0.19	3.00±0.02a	3.00±0.02a	2.66±0.02	11.33±0.57ab
	125	3.00±0.19	3.00±0.02a	3.00±0.02a	2.66±0.02	11.66±0.57a
CuSo4	75	2.33±0.19	2.66±0.02b	2.33±0.02ab	2.00±0.02	9.33±0.57bc
	100	2.66±0.19	2.66±0.02b	2.33±0.02ab	2.33±0.02	10.00±0.57ab
	125	2.66±0.19	2.66±0.02b	2.66±0.02a	2.33±0.02	10.33±0.57ab

^{a,b} Means with different superscript in the same column are significantly different at (P<0.05).
CuSo4 = Copper sulfate; Cu-NP = Copper nanoparticles.

Table 6. Effects of dietary copper supplementation on fatty acid composition in broiler chickens.

Items	Levels	Myristic	Stearic	Palmitic	Palmitoleic	Oleic	Linoleic	Linolenic	Arachidonic	TSFAs	TMUFAs	TPUFAs
	(mg/kg)	14:0	18:0	16:0	16:1	18:1	18:2	18:3	20:4	-----	-----	-----
Sources	Cu -NP	47.3±4.0b	469±12.1	1174±21.0a	400±6.0a	1533±7.9a	469±12.1b	165±6.9a	151±4.8a	1714±36.0a	1934±15.8a	1136±19.1a
	Cu sulfate	70.6±4.0a	471±12.1	1230±21.0a	352±6.0b	1474±7.9b	617±12.1a	132±6.9b	117±4.8b	1806±36.0b	1838±15.8b	974±19.1b
Copper level	0	97.0±4.9a	621±8.6a	1386±12a	316±10.0c	1429±12c	584±9.9c	114±6.8c	110±7.5b	2152±20a	1745±22c	883.0±35c
	75	73.5±4.9b	508±8.6b	1273±12b	357±10.0b	1475±12b	620±9.9b	125±6.8bc	118±7.5b	1889±20b	1832±22b	962.5±35bc
	100	58.0±4.9bc	455±8.6c	1205±12c	374±10.0ab	1507±12ab	637±9.9ab	147±6.8b	134±7.5ab	1748±20c	1898±22ab	1064±35ab
	125	45.5±4.9c	447±8.6c	1128±12c	399±10.0a	1529±12a	665±9.9b	173±6.8c	149±7.5a	1644±20d	1928±22a	1139±35a
Control	0	97.0±0.57a	621±7.6a	1386±15.3a	316±7.1g	1429±14.2f	584±2.7g	114±2.7f	110±2.7f	2152±22a	1745±18.2c	883.0±12.4c
	75	61.0±0.57d	493±7.6c	1246±15.3c	383±7.1c	1504±14.2c	643±2.7c	139±2.7d	128±2.7c	1830±22c	1887±18.2b	1024±12.4c
Cu -NPs	100	45.0±0.57f	439±7.6f	1185±15.3e	399±7.1b	1541±14.2b	659±2.7b	167±2.7b	152±2.7b	1693±22e	1940±18.2a	1159±12.4b
	125	36.0±0.57g	475±7.6d	1092±15.3g	419±7.1a	1556±14.2a	691±2.7a	189±2.7a	173±2.7a	1621±22g	1975±18.2a	1225±12.4a
CuSo4	75	86.0±0.57b	524±7.6b	1301±15.3b	331±7.1f	1446±14.2e	598±2.7f	112±2.7g	109±2.7f	1949±22b	1777±18.2c	901±12.4e
	100	71.0±0.57c	471±7.6e	1225±15.3d	349±7.1e	1474±14.2d	615±2.7f	128±2.7e	117±2.7e	1803±22d	1856±18.2b	970±12.4d
	125	55.0±0.57e	419±7.6g	1164±15.3f	378±7.1d	1503±14.2c	639±2.7d	158±2.7c	125±2.7d	1667±22f	1881±18.2b	1053±12.4c

^{a-f} Means within the same column without common superscripts are significantly different ($P < 0.05$).

TSFAs = Total saturated fatty acid; TMUFAs = Total mono-unsaturated fatty acid; TPUFAs = Total poly-unsaturated fatty acid.

References

- Amaral, G.V., Silva, E.K., Costa, A.L.R., Alvarenga, V.O., Cavalcanti, R.N., Esmerino, E.A., Guimarães, J.T., Freitas, M.Q., Sant'Ana, A.S., Cunha, R.L. and Moraes, J., 2018. Whey-grape juice drink processed by supercritical carbon dioxide technology: Physical properties and sensory acceptance. *Lwt*, 92 : 80-86.
- American Oil Chemists Society "AOCS"., 1993. Official Methods and Recommendation Practices of the American Oil Chemists Society. 4th Ed. Published by American Oil Chemists Society, 1608, Broad Moor drive, Champaign, USA.
- Association of Official Analytical Chemists "AOAC"., 2005. Official Methods of Analysis. 14th Ed., Horwitz, W.; D. (Editor), Academic Press, Washington D.C, USA.
- Aura, A.; Forssell, P.; Mustranta, A. and Poutanen, K., 1995. Transesterification of soy lecithin by lipase and phospholipase. *J. Amer. Oil Chem. Soc.*, 72 (11): 1375-1379.
- Bao, Y.M. and Choct, M., 2009. Trace mineral nutrition for broiler chickens and prospects of application of organically complexed trace minerals: a review. *Animal Production Science*, 49(4) : 269-282.
- Bianchi, M., M. Petracci and C. Cavani., 2006. "The influence of genotype, market live weight, transportation, and holding conditions prior to slaughter on broiler breast meat color." *Poultry science* 85(1): 123-128.
- Boni, I., H. Nurul and I. Noryati., 2010. "Comparison of meat quality characteristics between young and spent quails." *Int. Food Res. J* 17(3): 661-666.
- Choi, I.H., Park, W.Y. and Kim, Y.J., 2010. Effects of dietary garlic powder and α -tocopherol supplementation on performance, serum cholesterol levels, and meat quality of chicken. *Poultry Science*, 89(8), pp.1724-1731.
- Dadgar, S., E. Lee, T. Leer, N. Burlingquette, H. Classen, T. Crowe and P. Shand., 2010. Effect of microclimate temperature during transportation of broiler chickens on quality of the pectoralis major muscle." *Poultry science* 89(5): 1033-1041.
- Duncan, David B. 1955. "Multiple Range and Multiple F Tests." *Biometrics* 11 (1):1-42. doi: 10.2307/3001478.
- El Basuini, M.F., El-Hais, A.M., Dawood, M.A., Abou-Zeid, A.E.S., EL-Damrawy, S.Z., Khalafalla, M.M.E.S., Koshio, S., Ishikawa, M. and Dossou, S., 2016. Effect of different levels of dietary copper nanoparticles and copper sulfate on growth performance, blood biochemical profiles, antioxidant status and immune response of red sea bream (*Pagrus major*). *Aquaculture*, 455 : 32-40.
- El-Basuini, M. F., El-Hais, A. M., Dawood, M. A. O., Abou-Zeid, A. E.-S., El- Damrawy, S. Z., Khalafalla, M. M. E.-S., Dossou, S., 2016. Effect of different levels of dietary copper nanoparticles and copper sulfate on growth performance, blood biochemical profiles, antioxidant status and immune response of red sea bream (*Pagrus major*). *Aquaculture*, 455, 32-40.
- Gonzales-Eguia, A., Fu, C.M., Lu, F.Y. and Lien, T.F., 2009. Effects of nanocopper on copper availability and nutrients digestibility, growth performance and serum traits of piglets. *Livestock Science*, 126(1-3):122-129.
- Guo, Q., Richert, B.T., Burgess, J.R., Webel, D.M., Orr, D.E., Blair, M., Fitzner, G.E., Hall, D.D., Grant, A.L. and Gerrard, D.E., 2006. Effects of dietary vitamin E and fat supplementation on pork quality. *Journal of animal science*, 84(11) : 3089-3099.
- Horwitz, W., 2000. Association of Official Analytical Chemists. Official methods of analysis of the Association of Official Analytical Chemist. Gaithersburg, MD, USA.
- Ibrahim, A.I., EL-Gendi, G.M., Nihad, A.A., Okasha, H.M. and El-Attrouny, M.M., 2022. Potential Effects of Different Dietary Copper Sources to Improve Productive Performance, Plasma Biochemical Parameters and Oxidative Response Activities of Broiler Chickens. *Journal of Animal and Poultry Production*, 13(8):111-118.
- Kim, B.C., Ryu, Y.C., Cho, Y.J. and Rhee, M.S., 2006. Influence of dietary α -tocopheryl acetate supplementation on cholesterol oxidation in retail packed chicken meat during refrigerated storage. *Bioscience, biotechnology, and biochemistry*, 70(4) :808-814.
- Kim, Y.J., Chang, Y.H. and Jeong, J.H., 2005. Changes of cholesterol and selenium levels, and fatty acid composition in broiler meat fed with garlic powder. *Food Science and Biotechnology*, 14(2), pp.207-211.
- Lee, I.-C., Ko, J.-W., Park, S.-H., Lim, J.-O., Shin, I.-S., Moon, C., Kim, J.-C. 2016. Comparative toxicity and biodistribution of copper nanoparticles and cupric ions in rats. *International Journal of Nanomedicine*, 11, 2883-2990.

- López-Ferrer S., Baucells M.D., Barroeta A.C., Grashorn M. A., 2001.** n-3 enrichment of chicken meat. 1. Use of very long-chain fatty acids in chicken diets and their influence on meat quality: fish oil. *Poultry Sci.*, 80,741-752.
- McDowell, L.R., 1992.** Minerals in animal and human nutrition. Academic Press Inc..
- Mead, G. 1987.** "Recommendation for a standardized method of sensory analysis for broilers." *World's poultry science journal*.
- Miroshnikov, S.A., Yausheva, E.V., Sizova, E.A., Miroshnikova, E.P. and Levahin, V.I., 2015.** Comparative assessment of effect of copper nano- and microparticles in chicken. *Oriental Journal of Chemistry*, 31(4) : 2327.
- Mroczek-Sosnowska, N., Łukasiewicz, M., Wnuk, A., Sawosz, E., Niemiec J., 2014.** Effect of copper nanoparticles and copper sulfate administered in ovo on copper content in breast muscle, liver and spleen of broiler chickens. *Annals of Warsaw University of Life Sciences-SGGW. Animal Science*, 53, 135–142.
- Mroczek-Sosnowska, N., Łukasiewicz, M., Wnuk, A., Sawosz, E., Niemiec, J., Skot, A., Jaworski, S. and Chwalibog, A., 2016.** In ovo administration of copper nanoparticles and copper sulfate positively influences chicken performance. *Journal of the Science of Food and Agriculture*, 96(9):3058-3062.
- Mroczek-Sosnowska, N., Sawosz, E., Vadalasetty, K.P., Łukasiewicz, M., Niemiec, J., Wierzbicki, M., Kutwin, M., Jaworski, S. and Chwalibog, A., 2015.** Nanoparticles of copper stimulate angiogenesis at systemic and molecular level. *International journal of molecular sciences*, 16(3) : 4838-4849..Joshua, P.P.
- Muralisankar, T., Bhavan, P.S., Radhakrishnan, S., Seenivasan, C. and Srinivasan, V., 2016.** The effect of copper nanoparticles supplementation on freshwater prawn *Macrobrachium rosenbergii* post larvae. *Journal of Trace Elements in Medicine and Biology*, 34: 39-49.
- Payvastegan, S., Farhoomand, P., & Delfani, N. (2013).** Growth performance, organ weights and blood parameters of broilers fed diets containing graded levels of dietary canola meal and supplemental copper. *The Journal of Poultry Science*, 50, 354–363.
- Pearson, D. 2006.** "Chemical Analysis of Foods." 11th Ed, Publishing Co.,.
- Pikul, J., D. E. Leszczynski and F. A. Kummerow., 1989.** "Evaluation of three modified TBA methods for measuring lipid oxidation in chicken meat." *Journal of Agricultural and Food Chemistry* 37(5): 1309-1313.
- Rahman, Z.U., Besbasi, F., Afan, A.M., Bengali, E.A., Zendah, M.I., Hilmy, M., Mukhtar, M., Jaspal, S. and Aslam, N., 2001.** Effects of copper supplement on haematological profiles and broiler meat composition. *Int J Agric Biol*, 3 : 203-205.
- SAS, Institute. 2004.** The SAS/STAT 9.1 User's Guide. Vol. 1–7. SAS Institute Cary, NC.
- Scott, A., Vadalasetty, K. P., Sawosz, E., Łukasiewicz, M., Vadalasetty, R. K. P., Jaworski, S., & Chwalibog, A. 2016.** Effect of copper nanoparticles and copper sulphate on metabolic rate and development of broiler embryos. *Animal Feed Science and Technology*, 220, 151–158
- Shahzad, M.N., Javed, M.T., Shabir, S., Irfan, M. and Hussain, R., 2012.** Effects of feeding urea and copper sulphate in different combinations on live body weight, carcass weight, percent weight to body weight of different organs and histopathological tissue changes in broilers. *Experimental and Toxicologic Pathology*, 64(3), pp.141-147.
- Sharif, M., Rahman, M.A.U., Ahmed, B., Abbas, R.Z. and Hassan, F.U., 2021.** Copper nanoparticles as growth promoter, antioxidant and anti-bacterial agents in poultry nutrition: prospects and future implications. *Biological Trace Element Research*, 199(10) : 3825-3836.
- Skrivan, M., Skrivanova, V., Marounek, M., Tumova, E., Wolf, J., 2000.** Influence of dietary fat source and copper supplementation on broiler performance, fatty acid profile of meat and depot fat, and on cholesterol content in meat. *British Poultry Science*, 41, 608–614.
- SKřivaovan, V., Skřivan, M., TůMOVÁ, E. and Ševčíková, S., 2011.** Influence of dietary vitamin E and copper on fatty acid profile and cholesterol content of raw and cooked broiler meat.
- Świątkiewicz, S., Arczewska-Włosek, A. and Jozefiak, D., 2014.** The efficacy of organic minerals in poultry nutrition: review and implications of recent studies. *World's Poultry Science Journal*, 70(3) : 475-486.
- Upadhaya, S. D., Lee, B. R., Kim, I. H., 2016.** Effects of ionised or chelated water-soluble mineral mixture supplementation on growth performance, nutrient digestibility, blood characteristics, meat quality and intestinal microbiota in broilers. *British Poultry Science*, 57, 251–256.
- Valli, C. and Balakrishnan, V., 2016.** Effect of in ovo supplementation of nano forms of zinc, copper, and selenium on post-hatch performance of broiler chicken. *Veterinary world*, 9(3): 287.
- Vogel, S. F., 1975.** Fatty acid composition of raw and processed meats. *Food Technol.* 29: 147-152.
- Wang, C., Wang, M. Q., Ye, S. S., Tao, W. J., Du, Y. J., 2011.** Effects of copper-loaded chitosan

nanoparticles on growth and immunity in broilers. Poultry Science, 90, 2223-2228.
Wang, C., Wang, M.Q., Ye, S.S., Tao, W.J. and Du, Y.J., 2011. Effects of copper-loaded chitosan nanoparticles on growth and immunity in broilers. Poultry science, 90(10) : 2223-2228.

Wang, M.Q., Du, Y.-J., Wang, C., Tao, W.-J., He, Y.-D., Li, H., 2012. Effects of copper-loaded chitosan nanoparticles on intestinal microflora and morphology in weaned piglets. Biological Trace Element Research, 149, 184–189.

تأثير التغذية علي اشكال ومستويات مختلفة من النحاس على خصائص الذبيحة وصفات جودة اللحم لدجاج التسمين

إبراهيم عبدالكريم إبراهيم 1 ، جعفر محمود الجندي 1 ، نهاد عبداللطيف علي 2 ، حمادة مجد عكاشة 1 ، محمود مصطفى الاطروني 1

1 قسم الإنتاج الحيواني – كلية الزراعة – جامعة بنها – مصر

2 قسم الإنتاج الحيواني – كلية الزراعة – جامعة القاسم الخضراء – العراق

يهدف هذا البحث إلى دراسة تأثير إضافة اشكال مختلفة من النحاس بمستويات 75 ، 100 و 125 مللجرام /كجم علف ، كبريتات النحاس الغير عضوية (CuSO₄) حبيبات النحاس النانوية (Cu-NP) عي كلا من : وزن الجسم الحي وخصائص الذبيحة وجودة اللحم لدجاج التسمين. تم استخدام عدد 420 كتكوت عمر يوم من سلالة دجاج روس 308 وزعت عشوائيا علي 7 معاملات تجريبية (60 طائرًا لكل منها). تم تغذية طيور المجموعة الأولى على عليقة اساس بدون أي اضافت (مجموعة مقارنة) ، وتم تغذية المجموعات الثانية والثالثة والرابعة علي العليقة الاساسية مضاف اليها كبريتات النحاس الغير عضوية بمستويات 75 ، 100 و 125 جزء في المليون ، وتم تغذية المجموعه الخامسة والسادسة والسابعة على العليقة الاساسية مضاف اليها حبيبات النحاس النانوية Cu-NP بمستويات 75 و 100 و 125 جزء في المليون. أظهرت النتائج أن الطيور التي تم تغذيتها على العليقة الأساسية المضاف إليها النحاس النانوية حسنت معنويا وزن جسم وخصائص الذبيحة وجودة اللحم بينما انخفضت نسبة الدهون مقارنة بمجموعة الكنترول. بينما ، أظهرت المجموعات المضاف اليها Cu-NP تحسناً ملحوظاً في خصائص الذبيحة وجودة اللحم مقارنة بالمجموعات الأخرى. أدت إضافة النحاس باشكاله المختلفة علي العليقة إلى انخفاض معنوي في نسبة الدهون في اللحم ، ودرجة الحموضة ، و TVN و TBA. من ناحية أخرى ، أدت إضافة النحاس بصوره المختلفه ايضا إلى زيادة معنوية في مستويات التقييم الحسي للحوم (اللون والمرونة) مقارنة بمجموعة الكنترول. أدت إضافة النحاس للعليقة إلى زيادة محتوى العضلات من الأحماض الدهنية الأحادية غير المشبعة (TMUFA) والأحماض الدهنية المتعددة غير المشبعة (TPUFA) وانخفض إجمالي الأحماض الدهنية المشبعة مقارنة بمجموعة الكنترول. في النهاية ، أدت إضافة حبيبات النحاس النانوية بمستوي 125 جزء في المليون لكل كيلو جرام عليقة إلى تحسين وزن الجسم الحي ونسبة النصافي والصفات الكيميائية للحوم في دجاج التسمين. أظهرت الطيور التي تم تغذيتها على Cu-NP علي نموًا خصائص كيميائية للحوم عن تلك المغذاه على النحاس غير العضوي (كبريتات النحاس).